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In-Vitro Antibacterial Activity of some Ganoderma Species: A Review

Asha Arora

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Abstract

The rising significance of personal health and wellbeing has spurred scientific interest in natural research on products. Numerous phytochemicals that are found naturally in plants, fruits, and vegetables have been discovered to have biological activity and are frequently hailed as being good for human health. In addition to current treatment approaches, herbal medications may be a safe and effective way to treat infectious infections. *Ganoderma* has long been used for the management of incessant infectious conditions such as diabetic foot ulcers, pneumonia, and chronic hepatitis. While there is little information on *Ganoderma*'s antiviral and antibacterial properties in humans, preliminary (*in vitro* and *in vivo*) research shows that the plant possesses a wide range of these properties. Furthermore, gram-positive and gram-negative bacteria are inhibited *in vitro* by antibacterial components found in *Ganoderma* species. The outcomes of preclinical (*in vitro*) and clinical investigations on the antibacterial and antifungal properties of *Ganoderma* species are brought to light in this review.

Pandya C¹, Arora A^{2,*}, and Mathur F³

¹ *Research Scholar, Department of Biotechnology, B N University, Udaipur (Rajasthan)*

² *Head, Department of Biotechnology, Udaipur (Rajasthan)*

³ *Research Scholar, Department of Botany, B N University, Udaipur (Rajasthan)*

*Corresponding Author, araudr@gmail.com

Diabetic Foot Ulcer

Diabetic foot ulcers (DFUs) are a frequent and possibly dangerous diabetic consequence^[1]. Out of the 537 million individuals with diabetes globally, 19 to 34% will experience a DFU at some point in their lives^[2]. It is an open sore or wound that typically develops on the foot's bottom or toes^[3]. It is caused by a number of diabetes-related conditions, some of which are difficult to cure. The main causes of DFU include a combination of neuropathy (nerve damage) and poor blood circulation, peripheral neuropathy (nerve damage), poor circulation, foot deformities (bunions, hammertoes),

calluses or corns, trauma or pressure on the feet, inadequate foot care and Obesity. Reduced blood circulation hinders the body's ability to heal and fight infections and increases the risk of developing diabetic foot ulcers.

Diabetic foot ulcer's symptoms and signs may include an open sore, redness, swelling, warmth, drainage or pus or signs of infection. It can lead to severe complications if left untreated, including cellulitis (skin infection), osteomyelitis (bone infection), gangrene (tissue death) and the potential need for limb amputation. About 20% of individuals with DFU will need lower-extremity amputations, either major (above the ankle), minor (below the ankle), or both. Ten percent of DFU patients will die within a year of their first diagnosis [4]. In some cases, systemic infections can occur, which can be life-threatening. Preventing DFU is a crucial aspect of diabetes care [5]. The healing process of a typical wound progresses through four stages: hemostasis, inflammation, proliferation, and remodelling. However, a diabetes patient's constant hyperglycaemia has an effect on a number of normal wound healing processes [6]. Wound healing in it can be a time-consuming process and the time it takes for complete healing can vary significantly from person to person. It is crucial for individuals with diabetes to seek prompt medical attention for any foot ulcer and to follow the recommendations of their healthcare team to optimize the chances of successful wound healing and prevent complications [7]. This involves maintaining good blood sugar control, regularly examining the feet for any signs of injury or pressure points, wearing comfortable and properly fitting shoes, practising proper foot hygiene, avoiding walking barefoot and seeking professional podiatric care. In more severe cases, surgical intervention may be necessary. DFU requires careful monitoring and management [8][9], often involving a healthcare team that may include podiatrists, wound care specialists and endocrinologists. Preventive care and early detection are crucial in avoiding the development of diabetic foot ulcers and their complications.

Diabetic foot ulcer is a polymicrobial infection harbouring different bacteria. Diabetic foot infections (DFI) are composed of a mixture of Gram positive (*Staphylococcus aureus*, *Bacillus subtilis*, *Streptococcus* and *Enterococcus spp*) and Gram negative (*Escherichia coli*, *Klebsiella pneumoniae*, *Morganella morganii*, *Enterobacter*, *Pseudomonas*, *Citrobacter*, *Salmonella* and *Proteus sp.*) bacteria [10]. Among these multi drug resistance is one of the challenges faced by the therapists. In addition to determining the lesion's origin, the analysis is conducted to determine whether the ulcer is neuropathic, ischemic, or neuro-ischemic and provide details about its size, depth, appearance, and location.

Ganoderma and its species

Ganoderma is one of the most popular and important wild medicinal mushrooms Genus that has been therapeutically used since thousands of years [11]. Numerous studies and research have confirmed the multidirectional biological activity of extracts from various *Ganoderma* spp. and isolated compounds. The following properties have been proven anti-diabetic [12], hypoglycaemia, anti-cancer [13], anti-inflammatory, anti-tumour [14], anti-oxidant [15], immunomodulatory, anti-viral, anti-bacterial [16], anticonvulsant, anti-fungal, antihypertensive, anti-atherosclerotic, anti-aging, anti-androgenic, anti-hepatotoxic, radical scavenging property, neuroprotection, sleep promotion, cholesterol synthesis inhibition, inhibition of lipid peroxidation/oxidative DNA damage, hepatoprotective properties, maintenance of gut health, prevention of obesity and stimulation of probiotics [17]. The most important groups of compounds found in *Ganoderma* spp. include triterpenes (*Ganoderma* triterpenes) and polysaccharides. Until now, more than 300 triterpenes and 200 polysaccharides

characterized by diverse chemical structures and biological activity have been isolated. It includes several species of mushrooms, each with its own characteristics and properties. Some of the most well-known *Ganoderma* species include:

Ganoderma applanatum

Also known as the Artist's Conk, this species has a woody appearance and is commonly found in North America and Europe. While it may not be as well-studied as some other *Ganoderma* species, it is used in traditional medicine for its potential medicinal properties ^[18]

Ganoderma boninense

G. boninense is a species of *Ganoderma* fungus which is a close relative of *G. lucidum* and it is native to various regions in Asia and has a more distinctive reddish-brown cap compared to other *Ganoderma* species. It is used in traditional medicine in certain Asian countries and is also under research for its potential health benefits.

Ganoderma lucidum

G. lucidum commonly known as Lingzhi or Reishi, and one of the most widely studied and revered species in traditional Chinese medicine. It has a glossy, varnished appearance and is known for its potential immunomodulatory, anti-inflammatory, antidiabetic and antioxidant properties. It is often used in herbal remedies and dietary supplements such as capsules, extracts, teas and dried slices ^[19].

Ganoderma tsugae (Hemlock Reishi)

This species, often called the Hemlock Reishi, is native to eastern North America and is closely related to *G. lucidum*. It is primarily found growing on hemlock trees (*Tsuga* species) and shares many of the potential health benefits of its close relative. *G. tsugae* is less common in commercial health products, but it can still be found in various forms such as dietary supplements, extracts and teas.

Ganoderma resinaceum

The Hemlock Varnish Shelf, is another species found in North America and Europe. It has a varnished cap and can grow on hardwood trees and not limited to a specific host tree and can be found on a variety of deciduous and coniferous trees. It is known for its potential medicinal properties, similar to other *Ganoderma* species.

Ganoderma australe

This species is found in Australia and parts of Southeast Asia. It shares some similarities with other *Ganoderma* mushrooms and is also used for its potential health-promoting properties.

These are just a few examples of *Ganoderma* species and there are many more within the genus such as *G. pfeifferi*, *G. oregonense*, *G. multipileum*, *G. adspersum*, *G. sessile*, *G. lipsiense*, *G. colossus*, *G. curtisii*, *G. lobatum*, *G. mbrekobenum*, *G. sinense*, *G. tornatum*, *G. tuberculosum*, *G. zonatum*, *G. miniatocinctum* and *G. weberianum* etc.

Hypoglycemic activity is demonstrated by numerous compounds present in the extracts of *G. lucidum*: polysaccharides, proteoglycans, proteins, and triterpenes. It is presumed that *G. lucidum* extracts may be an alternative adjuvant treatment for diabetes. The mechanism of action of polysaccharides is by increasing insulin levels and lowering blood glucose levels. A study on mice with type 2 diabetes showed that *Ganoderma spp.* is effective in regulating blood glucose levels and has a positive effect on the lipid profile; therefore, it is considered a good candidate in the treatment of type 2 diabetes with comorbid metabolic disorders. Ganoderan A and B isolated from an aqueous extract of *G. lucidum* showed anti-glycaemic properties. The Ling-Zhi-8 protein is effective in type 1 diabetes due to its immunomodulatory properties [20]. Ganodermin is a protein with antifungal activity isolated from *G. lucidum*. It inhibits the growth of *Botrytis cinerea*, *Fusarium oxysporum*, and *Physalosporapiricola*.

Exopolymers of *G. applanatum* noticeably decrease blood glucose levels. Ergosterol peroxide isolated from *G. applanatum* can inhibit aldose reductase, an enzyme which is involved in the development of diabetic complications (neuropathy and retinopathy); therefore, reductase inhibitors may be useful in their prevention. The latest reports have indicated the hypoglycaemic effect of extracts from two other species: *G. pfeifferi* and *G. resinaceum*. The aqueous extract of *G. resinaceum* led to a slight decrease in glycemia in alloxan-induced diabetic rats.

Ethanol extracts obtained from mycelium *G. applanatum*, *G. carnosum* and *G. lucidum* also demonstrated antifungal activity. They were effective against *Acremonium strictum*, *Aspergillus glaucus*, *Aspergillus flavus*, *Aspergillus fumigatus*, *Aspergillus nidulans*, *A. niger*, *Aspergillus terreus* and *Trichoderma viride*.

Antimicrobial activity

Genus *Ganoderma* has antimicrobial components that stop the growth of fungi, viruses and both gram-positive and gram-negative bacteria are inhibited from growing. Multidrug resistance (MDR) bacterial and fungal infections may be successfully treated with natural antimicrobial substances derived from a wide variety of medicinal plants [21]. *Ganoderma* extracts contain substances with antibacterial properties, which are caused by phytochemicals synthesised in the plant's secondary metabolism [22][23]. According to the World Health Organisation (WHO), medicinal plants would be the best source to obtain a range of medications. Plants include a number of secondary metabolites that have been shown to have antibacterial effects in vitro, including tannins, alkaloids, phenolic compounds and flavonoids [24][25].

Ganoderma species, contain bioactive compounds such as polysaccharides, triterpenes and peptides, with demonstrated antibacterial properties. These compounds exhibit a broad-spectrum antimicrobial activity, making them effective against a range of bacterial strains. Studies have indicated that extracts from *Ganoderma* species can inhibit the growth and multiplication of pathogenic bacteria commonly found in diabetic foot ulcers. This includes both Gram-positive (*Staphylococcus aureus*, *Enterococcus faecalis*, *Bacillus subtilis*, *Enterococcus faecium* and *Streptococcus pneumoniae*

etc) and Gram-negative (*Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *salmonella typhimurium* and *Proteus vulgaris* etc) bacteria. Bacterial biofilms are often implicated in chronic wound infections, including diabetic foot ulcers. *Ganoderma* extracts have demonstrated the ability to disrupt biofilms formed by pathogenic bacteria, making it easier for the immune system and antimicrobial agents to target and eliminate the bacteria.

There are many different methods are used to extract different bioactive metabolite from *Ganoderma*. Some of the methods which we are discussing here, are being used in today's time:

Numerous extracts obtained from *Ganoderma* with varying polarity (from non-polar to polar) have been utilised in scientific studies to explore the wide range of metabolites, including polysaccharides, triterpenoids, and phenols [26], alkaloids etc., present within this fungus. These extracts possess distinct chemical profiles, enabling targeted exploration of specific metabolite classes and their associated biological activities [27].

AgNO₃ was reduced to produce silver nanoparticles via a green process mediated by mycelial extracts of *G. lucidum* have improved stability and excellent dispersion in an aqueous solution, (AgNPs). Compared to other metal NPs, AgNPs have more antibacterial activity against multidrug-resistant bacteria. Gram-positive and Gram-negative bacterial and yeast strains were used to assess the synthetic nanoparticles' antibacterial efficacy. Because the pathogens were inhibited in their multiplication, the environment and public health were less at danger due to the effectiveness of the silver nanoparticles [28].

Using an environmentally friendly process, copper oxide nanoparticles (CuONPs) were created from the supernatant and extract of the fungus *G. sessile*, and their antibacterial and biocompatibility characteristics were identified. CuONPs showed antimicrobial activity against *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*. Further research might be done on the CuONPs created from the fungus *G. sessile* extract in order to see whether they are effective in treating superficial infectious illnesses. [29].

A green method was used to create new biogenic silver (AgNPs) and gold nanoparticles (AuNPs) using *G. lucidum* extract. *B. subtilis* showed the greatest growth suppression activity of GL-AgNPs, followed by *B. cereus*, *P. aeruginosa*, *E. coli*, and *S. aureus* [30].

Zinc oxide (ZnO) nanoparticles with different concentrations of *G. lucidum* extract have been biologically produced, and their optical, morphological, structural and elemental properties have been identified. The use of green generated ZnO with *G. lucidum* extracts as a nanonutrient is first documented in the study by Sedefogluet *al.*, 2022 [31].

Novel anti-biofilm strategies must be designed to include natural bio products instead of common antibiotics. Mushrooms are a nutritionally functional foods and a source of pharmaceuticals functions such as antidiabetic, antitumor, immunomodulating, antioxidant, cardiovascular, anti-hypercholesterolemia, antimicrobial. *Ganoderma*, especially *G. lucidum* have a notable activity against biofilms [32].

Table 1.

Species	Bacteria	Remark	Reference
<i>Ganoderma sessile</i>	<i>Staphylococcus aureus</i>	CuONPs-S (Copper oxide nanoparticles- <i>S. aureus</i>) were more cytotoxic to kidney cells and macrophages, and the hepatocytes	[33] [34] [35]
	<i>Escherichia coli</i>		
	<i>Pseudomonas aeruginosa</i>	CuONPs-E (Copper oxide nanoparticles- <i>E. coli</i>) were less cytotoxic to kidney cells and macrophages, and the hepatocytes	
<i>Ganoderma boninense</i>	<i>Staphylococcus aureus</i>	The highest antibacterial activity was observed in chloroform-extracted GBMA (<i>G. boninense</i> media agar) against all tested bacteria	[36] [37] [38]
	<i>Streptococcus pyogenes</i>		
	<i>Pseudomonas aeruginosa</i>	Methanol-extracted GBMA exhibited higher and broader ranges of antibacterial activity against <i>S. aureus</i>	
	<i>Klebsiella pneumonia</i>		
	<i>Escherichia coli</i>	Methanol and acetone extracted GBFB (extract of <i>G. boninense</i> fruiting bodies) and GBMA demonstrated lower antibacterial activity than chloroform extracted GBMA GBMB (<i>G. boninense</i> media broth) did not exhibit any antibacterial activity against <i>S. aureus</i> , MRSA, and <i>K. Pneumonia</i>	
<i>Ganoderma lucidum</i>	<i>Acidovorax avenae</i>	Only the methanol and water extracts showed inhibition of all the phytopathogens tested. <i>Erwinia carotovorasubsp. carotovora</i> and <i>P. syringae pv. phaseolicola</i> were the most inhibited, whereas <i>P. syringae pv. syringae</i> was least inhibited Culture fluids of <i>G. lucidum</i> , inhibited both gram-positive and gram-negative plant pathogenic bacteria	[39] [40] [41]
	<i>Agrobacterium rhizogenes</i>		
	<i>Agrobacterium tumefaciens</i>		
	<i>Brenneria quercina</i>		
	<i>Burkholderia cepacian</i>		
	<i>Erwinia carotovora</i>		
	<i>Pseudomonas fluorescens</i>		
	<i>Pseudomonas syringae pv. Syringae</i>		
	<i>Rathayibacter tritici</i>		
	<i>Xanthomonas campestris pv. Campestris</i>		
<i>Ganoderma lipsiense</i>	<i>Staphylococcus aureus</i>	Crude extract of <i>G. lipsiense</i> and their fractions dichloromethane and ethyl acetate showed antibacterial activities against <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> at 500 µg mL ⁻¹ . The production of ergosta-6,22-diene-3β,5α,8α-triol by <i>G. lipsiense</i> showed antiparasitic activity against <i>Giardia duodenalis</i> trophozoites	[42] [43] [44]
	<i>Pseudomonas aeruginosa</i>		
<i>Ganoderma austral</i>	<i>Escherichia coli</i>	The nucleotide sequences obtained from all <i>Ganoderma</i> strains in this study were deposited in GenBank. <i>G. austral</i> and <i>G. applanatum</i> extracts showed anti-proliferative activity against the tumor cell at concentration lower than 50 µg/mL <i>G. curtisii</i> and <i>G. lucidum</i> showed anti-proliferative activity and remarkable inhibition values for HeLa cell line <i>G. curtisii</i> show such growth inhibitory mechanisms in tumor cells	[45]
<i>Ganoderma applanatum</i>			
<i>Ganoderma curtisii</i>			
<i>Ganoderma lobatum</i>			
<i>Ganoderma lucidum</i>			
<i>Ganoderma oregonense</i>			
<i>Ganoderma resinaceum</i>			
	<i>Escherichia coli</i>	Triterpenoids, (ganoderic acids) responsible for its antitumor activity.	
	<i>Salmonella typhi</i>		
	<i>Staphylococcus aureus</i>		

Ganoderma lucidum	(WNSA)	β-Glucans (polysaccharide) develops protective inflammatory responses that prevent infections by pathogens, including infections by corona viruses. <i>G. lucidum</i> can be used in the treatment of COVID -19 infections.	[46][47][48]
	<i>Streptococcus pyogenes</i>		
	<i>Bacillus subtilis</i>		
	<i>Enterobacter aerogenes</i>		
	<i>Corynebacterium diphtheria</i>		
	<i>Pseudomonas aeruginosa</i>		
Ganoderma mbrekobenum	<i>Bacillus subtilis</i>	The higher antibacterial activity produced by methanol extract was against all tested pathogens	[49][50][51]
	<i>Bacillus cereus</i>		
	<i>Fusarium oxysporum</i> (fungus)		
Ganoderma multiplicatum Ganoderma sinense	<i>Staphylococcus aureus</i>	The compounds ganosinensin B and ganosinoside A, present in extracts of both <i>Ganoderma</i> species showed strong antibacterial activity against <i>S. Aureus</i>	[52][53][54][55]
Ganoderma tsugae	<i>Staphylococcus aureus</i>	Triterpenoids present in <i>G. tsugae</i> extract showed strong antibacterial activity against all tested bacteria	[56][57]
	<i>Bacillus subtilis</i>		
	<i>Escherichia coli</i>		
Ganoderma tornatum Ganoderma tuberculosum	<i>Clavibacter michiganensis</i>	<i>G.tuberculosum</i> and <i>G.martinicense</i> had the best antioxidant and antibacterial activity	[58][59][60]
Ganoderma lucidum	<i>Staphylococcus aureus</i>	AgNPs (silver nanoparticles) were successfully synthesized from <i>G. lucidum</i> crude extracts and these colloidal AgNPs demonstrated an extraordinary antimicrobial activity against all tested pathogens	[61][62]
	<i>Escherichia coli</i>		
	<i>Pseudomonas aeruginosa</i>		
	<i>Salmonella enteric</i>		
	<i>Candida albicans</i>		
Ganoderma lucidum	<i>Enterococcus faecalis</i>	Methanolic extract of GL exhibited higher antibacterial activity against <i>E. coli</i> and <i>P. aeruginosa</i>	[63][64][65][66]
	<i>Staphylococcus aureus</i>		
	<i>Escherichia coli</i>		
	<i>Pseudomonas aeruginosa</i>		
	<i>B. subtilis</i>		
Ganoderma oerstedii, Ganoderma weberianum Ganoderma subincrustatum	<i>Staphylococcus aureus</i> <i>Escherichia coli</i>	Antibacterial activity was weak against <i>S. aureus</i> All tested <i>Ganoderma</i> mushrooms have medicinal potential such as anti-inflammatory and anti-proliferative.	[67][68][69][70]
Ganoderma neojaponicum Ganoderma lucidum	<i>Salmonella typhimurium</i>	compared to <i>G. lucidum</i> , <i>G. neo-japonicum</i> showed remarkable antibacterial and antioxidant properties	[71][72]
	<i>Salmonella Enteritidis</i>		
	<i>Escherichia coli</i>		

Ganoderma applanatum	<i>Escherichia coli</i>	The synthesized AgNPs from methanolic extract of <i>G. applanatum</i> exhibit high <u>antioxidant capacity</u> , in vitro <u>antibacterial activity</u> against <i>S. aureus</i> and <i>E. coli</i> and in vivo antifungal activity <i>G. applanatum</i> can be efficiently used in synthesis of AgNPs with potent antimicrobial properties, which can be used for both clinical and <u>agrochemical</u> purposes.	[73]
	<i>Staphylococcus aureus</i>		
	Fungus- <i>Botrytis cinerea</i>		
	<i>Colletotrichum gloeosporioides</i>		
Ganoderma atrum	<i>Escherichia coli</i>	<i>G. atrum</i> is a popular remedy to treat conditions such as chronic hepatitis, hypertension, cancer, hyperlipemia, bronchitis, atherosclerosis and diabetes The antibacterial activity of each group was shown as <i>G. atrum</i> sterol components > ergosterol > ergosterol ester.	[74][75][76]
	<i>Salmonella enteric</i>		
	<i>Staphylococcus aureus</i>		
	<i>Shigella sonnei</i>		
	<i>Listeria monocytogenes</i>		
Ganoderma applanatum Ganoderma lucidum	<i>Pseudomonas fluorescens</i> <i>Bacillus subtilis</i> <i>Staphylococcus epidermidis</i> <i>Micrococcus luteus</i> <i>Pseudomonas aeruginosa</i>	Water and methanol extracts of both <i>Ganoderma</i> exhibited strong antibiotic activity against all bacterial strains tested.	[77][78]
Ganoderma lucidum	<i>Lepidium sativum</i> (garden cress)	first report of green synthesized ZnO with <i>G. lucidum</i> extracts The effect of extract concentration on various properties of ZnO Nano Particles Nano nutrient effect of ZnO NPs on <i>Lepidium sativum</i>	[79][80][81]
Ganoderma applanatum, Ganoderma lucidum Ganoderma pfeifferi Ganoderma resinaceum	<i>Pseudomonas aeruginosa</i> <i>Staphylococcus aureus</i> <i>Escherichia coli</i> <i>Aspergillus niger</i>	Chloroform extract of <i>G. resinaceum</i> expressed the most potent antibacterial activity against <i>P. aeruginosa</i> Aqueous extract of <i>G. pfeifferi</i> expressed the most potent antibacterial activity against both <i>E. coli</i> and <i>S. aureus</i> Ethanol extracts of <i>G. pfeifferi</i> and <i>G. resinaceum</i> were the most effective against <i>A. Niger</i>	[82][83][84]
Ganoderma boninense	<i>Staphylococcus aureus</i> (MRSA)	<i>G. boninense</i> extract induces irreversible damage to the cell membrane of MRSA, thus causing cellular lysis and death	[85][86][87]
Ganoderma multipileum	<i>Klebsiella pneumoniae</i> <i>Staphylococcus aureus</i> <i>Escherichia coli</i> <i>Pseudomonas aeruginosa</i> Fungus- <i>Aspergillus flavus</i>	The ZnO nanoparticles synthesized from <i>G. multipileum</i> showed a strong antibacterial effect against gram-positive (<i>K. pneumoniae</i> and <i>S. aureus</i>) and gram-negative (<i>E. Coli</i> and <i>P. aeruginosa</i>) bacteria The ZnO nanoparticles also showed a high antifungal effect against all different fungus	[88][89][90]

	<i>Aspergillus niger</i>		
	<i>Aspergillus fumigatus</i>		
	<i>Fusarium solani</i>		
	<i>Mucor species</i>		
<i>Ganoderma cochlear</i>	<i>Staphylococcus aureus</i>	exhibit potent inhibitory activity against <i>S. Aureus</i>	[91][92][93]
<i>Ganoderma pfeifferi</i>	<i>Staphylococcus aureus</i>	Extracts of the fruiting bodies of the mushroom exhibited antibacterial activity against all tested bacteria	[94][95][96]
	<i>Bacillus subtilis</i>		
	<i>Escherichia coli</i>		
<i>Ganoderma austral</i>	<i>Escherichia coli</i>	The extracts of <i>G. curtisi</i> inhibited the growth of <i>S. aureus</i> Ganoderic acids, phenolic compounds may also be responsible for antibacterial inhibition and antioxidant activity of all tested <i>Ganoderma</i> species	[97][98]
	<i>Staphylococcus aureus</i>		
<i>Ganoderma applanatum</i>	<i>Pseudomonas aeruginosa</i>		
<i>Ganoderma colossus</i>			
<i>Ganoderma curtisii</i>			
<i>Ganoderma lobatum</i>	<i>Enterococcus faecalis</i>		
<i>Ganoderma oregonense</i>			
<i>Ganoderma resinaceum</i>			
<i>Ganoderma sessile</i>	<i>Escherichia coli</i>	A very low concentration of silver nanoparticles produced from the extract of <i>G. sessile</i> is required for the bacterial inhibition	[99][100][101]
	<i>Staphylococcus aureus</i>		
	<i>Pseudomonas aeruginosa</i>		

Antimicrobial Activity of *Ganoderma* Species against Gram Positive Bacteria

Fungi are particularly appealing for nano particle synthesis because they release huge amounts of enzymes and metabolites and are facile to manage in the laboratory. Copper and copper oxide NP synthesis has recently attracted attention, as recent research demonstrated that they are advantageous for biomedical applications due to their antibacterial, anticancer, antidiabetic and antioxidant characteristics [61]. Micro, quasi-spherical nanoparticles (NPs) with atypical size of 4.5 ± 1.9 nm and 5.2 ± 2.1 nm were obtained from the resulting supernatant and extract of the fungi *Ganoderma sessile*. CuONPs displayed antibacterial efficacy against *Staphylococcus aureus* (*S. aureus*). The half-maximal inhibitory concentration (IC50) value for *S. aureus*, was 10.2 µg/mL. When bacteria were subjected to CuONPs, their ultrastructural examination confirmed that tiny CuONPs were present throughout the bacterial cells [33]. The mycelial extract of *Ganoderma boninense* was found to be effective in producing secondary metabolites with antibacterial efficacy against *S. aureus* and *S. pyogenes*. Methanol-extracted GBMA exhibited higher and broader ranges of antibacterial activity against *S. aureus* [34]. Additionally, the minimum inhibitory concentration (MIC) of *Ganoderma sinense* and *Ganoderma multiplicatum* extracts demonstrated bactericidal activity against *S. aureus* [52]. Gram-positive and gram-negative plant pathogenic bacteria were suppressed by *G. lucidum* culture fluids. *Rathayibacter tritici* which is a plant pathogen of wheat was inhibited by extracts of *G. lucidum* [39]. The biochemical function of the distillates derived from the in vitro culture of Mexican strains of *Ganoderma* viz. *Ganoderma austral*, *Ganoderma applanatum*, *Ganoderma colossus*, *Ganoderma*

curtisii, *Ganoderma lobatum*, *Ganoderma lucidum*, *Ganoderma oregonense* and *Ganoderma resinaceum*. The growth of gram-positive *S. aureus* was suppressed by extracts of three strains of *G. curtisii* that exhibited anti-proliferative activity [45]. Research on *G. lucidum* extracts was evaluated on gram-positive and methicillin-resistant bacteria, including *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* (MRSA). *Streptococcus pyogenes* ($p = 0.05$) and *Staphylococcus aureus* (MRSA) ($p = 0.022$) were significantly inhibited by hexane extract of *Ganoderma lucidum*. *Streptococcus pyogenes* was the most sensitive microorganism [46]. There are scanty researches on the physiological functions of *G. mbrekobenum*'s fruiting bodies. Methanol extract of *G. mbrekobenum* demonstrated greater antibacterial activity against *Bacillus cereus* and *Bacillus subtilis*, measuring 14.13 ± 0.12 mm and 13.03 ± 0.12 mm, respectively. The majority of the test bacterial strains were resistant to the antibacterial effect of the aqueous extracts [49]. The antibacterial activity of *Ganoderma* strains viz. *G. tuberculosum*, *G. tornatum* and *G. weberianum* against *Clavibacter michiganensis*, which causes tomato canker, is highlighted within the concentration range 31.5 to 1000 $\mu\text{g/mL}$ is noteworthy [58]. Bacallao-Escudero et al. (2023) investigated the antibacterial activity of ethanolic extracts of *Ganoderma oerstedii*, *G. weberianum* and *G. subincrustatum* fruiting bodies against *Staphylococcus aureus* and *Escherichia coli* using the broth microdilution method. There was minimal antibacterial activity ($\text{MIC}_{50} > 10$ mg/mL) against *S. aureus* [67]. The strongest antibacterial activities against the studied pathogens were demonstrated by the "green synthesis" of silver nanoparticles (AgNPs) from *Ganoderma applanatum*. High antioxidant capacity, in vitro antibacterial activity against *Escherichia coli* and *Staphylococcus aureus*, and in vivo antifungal capabilities against *Colletotrichum gloeosporioides* and *Botrytis cinerea* are all displayed by the synthesised Ag nano particles [73]. ZnO nanoparticles synthesised from *Ganoderma multipileum* displayed significant antibacterial activity against gram-positive bacteria, including *Staphylococcus aureus* and *Klebsilla pneumonia* [88]. *G. boninense* is identified as an oil palm pathogen, although there is scanty information about its biological activity. In broth microdilution experiments, high susceptibility was reported in methicillin-resistant *Staphylococcus aureus* (MRSA) in the elute fraction, with a MIC value of 0.078 mg mL⁻¹. According to the findings, *G. boninense* extract causes irreversible damage to MRSA cell membranes, resulting in cellular lysis and death [85].

Antimicrobial Activity of *Ganoderma* Species against Gram Negative Bacteria

Gram-negative bacilli are the most common bacterial pathogens and are often resistant to medicinal products. Monitoring for antimicrobial resistance in this population is critical since resistance has been linked to increased morbidity and mortality. CuONPs extracted from the fungi *Ganoderma sessile* displayed antibacterial efficacy against gram-negative bacteria like *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Escherichia coli* (*E. coli*). The half-maximal inhibitory concentration (IC_{50}) values were 4.1 $\mu\text{g/mL}$ and 8.5 $\mu\text{g/mL}$, respectively [33]. The mycelial extract of *G. boninense* was found to be effective in producing secondary metabolites with antibacterial activity against *E. coli* and *P. aeruginosa* and *K. pneumonia* [36]. It was observed via liquid-liquid extractions (LLE) that mycelia extraction using a 1:1:1 combination of chloroform, methanol, and water was better at detecting antibacterial activity with the highest concentrations of antibacterial substances. Research have been conducted to isolate and distinguish the antibacterial activity of *Ganoderma lucidum* culture fluids against several gram-negative plant pathogenic bacteria like *Acidovorax avenae*, *Agrobacterium*

rhizogenes, *Agrobacterium tumefaciens*, *Brenneria quercina*, *Burkholderia cepacia*, *Erwinia carotovora*, *Pseudomonas fluorescens*, *Xanthomonas campestris*. Nearly all of the studied bacteria were unable to grow in the freshly obtained culture fluids of *Ganoderma lucidum*. *Erwinia carotovora subsp. carotovora* and *P. syringae pv. phaseolicola* were the most inhibited bacteria against the extracts of *G. lucidum*. *Pseudomonas fluorescens* and *Burkholderia cepacia* did not exhibit any form of activity. There was a little inhibition seen with *Brenneria quercina* [39]. The remarkable antibacterial and anticancer activities of *Ganoderma lucidum* methanolic extract (GLME) have garnered significant interest. Screening the extract's antibacterial properties against four strains of both Gram-positive and Gram-negative bacteria revealed that it had more antibacterial properties against *E. coli* bacteria than streptomycin, resulting in a zone of inhibition measuring 44 ± 0.09 mm [66]. Furthermore, colloidal AgNPs derived from *G. lucidum* displayed exceptional antibacterial efficacy against gram negative *Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella enterica* and *Candida albicans* in biological assays, with IC50 values of 17.06, 1.32, 54.69 and 27.78 g/mL, respectively [61]. Crude extracts of *G. lipiense*, along with their dichloromethane (DCMf) and ethyl acetate (Eaf) derivatives, demonstrated antibacterial activity at 500 $\mu\text{g mL}^{-1}$ against *Pseudomonas aeruginosa* and *Staphylococcus aureus* [42]. El-Dein et. Al. (2023) demonstrated that the methanolic extract of *G. mbrekobenum* exhibited the strongest antifungal activity against *F. oxysporum* and *F. oxysporum f. sp. Lycopersici* [49]. Experiments comparing the bioactivities of various *Ganoderma* species have been carried out. *G. lucidum* (GL) and *G. neo-japonicum* (GnJ) were extracted using hot water and their antimicrobial properties were contrasted. The pathogens *Salmonella typhimurium*, *Salmonella Enteritidis* and *Escherichia coli* had minimum inhibitory concentrations (MICs) of 1.25 mg/mL to 2.5 mg/mL for GL and 2.5 mg/mL to 5 mg/mL for GnJ. SEM demonstrated that the two extracts worked by lysing the cells and shrinking the pathogens' cell walls [71]. Investigations were conducted to evaluate the antimicrobial potential of the autochthonous *Ganoderma* species (*G. resinaceum*, *G. pfeifferi*, *G. lucidum* and *G. applanatum*). CHCl₃ extract of *G. resinaceum* had the strongest antibacterial activity against *P. aeruginosa*, EtOH extracts of *G. pfeifferi* and *G. resinaceum* were shown to have the strongest antibacterial activity against *A. niger*, whereas *G. pfeifferi* exhibited the maximum antibacterial activity against both *E. coli* and *S. aureus* [82]. ZnO nanoparticles that were isolated from *Ganoderma multipileum* had substantial antibacterial activity against gram-negative bacteria, such as *Escherichia coli* and *Pseudomonas aeruginosa* [88]. AgNPs were produced using *Ganoderma sessile* extracts and supernatants and their *in vitro* antibacterial efficacy against *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* was assessed. Once the minimum inhibitory concentration (MIC) was established, the AgNPs demonstrated antibacterial activity against all employed bacteria. The MIC ranged from 1.26 to 5.0 $\mu\text{g/mL}$, contingent upon the type of bacterium [99].

Conclusion

Ganoderma species are also known for their wound healing properties. By accelerating the wound healing process, they can indirectly contribute to reducing the risk of infection in diabetic foot ulcers and other wounds. *Ganoderma* species have immunomodulatory effects that can enhance the body's immune response. This can help the immune system combat bacterial infections more effectively, especially in individuals with diabetes, who may have compromised immune function. Inflammation is a key component of the body's response to bacterial infections. *Ganoderma*'s anti-inflammatory

properties can help reduce inflammation associated with diabetic foot ulcers and the corresponding infections.

This potential antimicrobial benefits of *Ganoderma* species for wound care in diabetic foot ulcers can be considered as complementary to standard medical treatment. Consulting with a healthcare professional is essential when using natural remedies like *Ganoderma*, as part of a comprehensive wound care plan. Furthermore, the quality and source of *Ganoderma* products can significantly impact their efficacy, so using reputable products is crucial.

Other References

- Shakeri F, Zaboli F, Fattahi E and Babavalian H. Effectiveness of Alginate Hydrogel Containing Ganoderma Polysaccharides on Wound Healing in Rat Model. *Trauma Monthly*. 2023; 28(1): 734-47.

References

1. [^]Chang M and Nguyen TT. Strategy for treatment of infected diabetic foot ulcers. *Acc. of chemical res*. 2021; 54(5): 1080-93.
2. [^]Armstrong DG, Boulton AJM and Bus SA. Diabetic Foot Ulcers and Their Recurrence. *N. Engl. J. Med*. 2017; 376(24): 2367-75.
3. [^]Monteiro-Soares M, Boyko EJ, Jeffcoate W, Mills JL, Russell D, Morbach S and Game F. Diabetic foot ulcer classifications: A critical review. *Diabetes Metab Res Rev*. 2020; 36 Suppl 1: e3272.
4. [^]Meloni M, Izzo V, Giurato L, Lázaro-Martínez JL and Uccioli L. Prevalence, clinical aspects and outcomes in a large cohort of persons with diabetic foot disease: comparison between neuropathic and ischemic ulcers. *J. Clin. Med*. 2020; 9:1780.
5. [^]Matoori S. (Topic Editor, ACS Pharmacology & Translational Science). Breakthrough Technologies in Diagnosis and Therapy of Chronic Wounds. *ACS Pharmacol and Translational Sci*. 2023; 6(6): 854-56.
6. [^]Liu Y, Liu Y, Deng J, Li W and Nie X. Fibroblast growth factor in diabetic foot ulcer: progress and therapeutic prospects. *Frontiers in endocrinol*. 2021; 12: 744868.
7. [^]Dixon D and Edmonds M. Managing diabetic foot ulcers: pharmacotherapy for wound healing. *Drugs*. 2021; 81(1): 29-56.
8. [^]Vileikyte L, Pouwer F and Gonzalez JS. Psychosocial research in the diabetic foot: are we making progress? *Diabetes Metab Res Rev*. 2020; 36(Suppl. 1): e3257
9. [^]Khunkaew S, Fernandez R and Sim J. Health-related quality of life among adults living with diabetic foot ulcers: a meta-analysis. *Qual. Life Res*. 2019; 28: 1413-27.
10. [^]Sadeghpour Heravi F, Zakrzewski M, Vickery KG, Armstrong D and Hu H. Bacterial diversity of diabetic foot ulcers: current status and future perspectives. *J. of clin. med*. 2019; 8(11):1935.
11. [^]Sun YF, Xing JH, He XL, Wu DM, Song CG, Liu S, Vlasak J, Gates G, Gibertoni TB and Cui BK. Species diversity, systematic revision and molecular phylogeny of Ganodermataceae (Polyporales, Basidiomycota) with an emphasis on

- Chinese collections. *Stud. Mycol.* 2022; 101: 287–415.
12. [^]Bhat ZA, Wani AH, War JM and Bhat MY. Major bioactive properties of *Ganoderma polysaccharides*: A review. *Asian J. of Pharmaceu. and Clin. Res.* 2021; 7:11-24.
 13. [^]Dora J and Hena VR. Antimicrobial activity of *G. lucidum* fruiting body extract from Himachal Pradesh. *Int. J. of Sci. Development and Res.* 2020; 5: 263-66.
 14. [^]Gong Z, Liu M, Liu H, Deng Z, Qin X, Nie J, Qiao Z, Zhu H and Zhong S. Structural features and in vitro antitumor activity of a water-extracted polysaccharide from *G. applanatum*. *New J. of Chem.* 2023; 47(28): 13205-17.
 15. [^]Huang P, Luo FJ, Ma YC, Wang SX, Huang J, Qin DD, Xue FF, Liu BY, Wu Q, Wang XL and Liu GQ. Dual antioxidant activity and the related mechanisms of a novel pentapeptide GLP4 from the fermented mycelia of *G. lingzhi*. *Food and Func.* 2022; 13(17): 9032-48.
 16. [^]Hu J, Li GF, Xu FM, Li Q, Lv T, Peng TF, Yin S, Gong W. Antibacterial lanostane triterpenoids from *G. tsugae*. *J. of Asian Nat. Products Res.* 2023; 4:1-7.
 17. [^]Wang L, Li JQ, Zhang J, Li ZM, Liu HG and Wang YZ. Traditional uses, chemical components and pharmacological activities of the genus *Ganoderma* P. Karst.: A review. *RSC Adv.* 2020; 10: 42084-097.
 18. [^]Luangharn T, Karunarathna SC, Dutta AK, Paloi S, Promputtha I, Hyde KD, Xu J and Mortimer PE. *Ganoderma* (Ganodermataceae, Basidiomycota) Species from the Greater Mekong Subregion. *J. Fungi.* 2021; 7: 819.
 19. [^]Fryssouli V, Zervakis GI, Polemis E and Typas MA. A global meta-analysis of ITS rDNA sequences from material belonging to the genus *Ganoderma* (Basidiomycota, Polyporales) including new data from selected taxa. *MycKeys.* 2020; 75: 71–143.
 20. [^]Cho JY, Sadiq NB, Kim JC, Lee B, Hamayun M, Lee TS, Kim HS, Park SH, Nho CW and Kim HY. Optimization of antioxidant, anti-diabetic and anti-inflammatory activities and ganoderic acid content of differentially dried *G. lucidum* using response surface methodology. *Food Chem.* 2021; 335:127645.
 21. [^]Savin S, Craciunescu O, Oancea A, Ilie D, Ciucan T, Antohi LS, Toma A, Nicolescu A, Deleanu C and Oancea F. Antioxidant, cytotoxic and antimicrobial activity of chitosan preparations extracted from *G. lucidum* mushroom. *Chem. and Biodiver.* 2020; 17(7): e2000175.
 22. [^]Thapa R, Maharjan R, Tamang P, Gautam P, Adhikari R and Maharjan S. Antimicrobial assessment and phytochemical screening of medicinal plants and *G. lucidum*. *Int. J. Appl. Sci. Biotechnol.* 2022; 10: 228-36.
 23. [^]Chan YS, Chong KP. Bioactive compounds of *G. boninense* inhibited methicillin-resistant *S. aureus* growth by affecting their cell membrane permeability and integrity. *Molecules.* 2022; 27(3): 838.
 24. [^]Ghosh S, Das S, Saha R, Acharya K. Studies of Antioxidant and Cytotoxic Activity in Ready-to-Drink Wild *Ganoderma* Teas: An In Vitro Approach. *Int. J. of Med. Mushrooms.* 2023; 25(11): 53-63.
 25. [^]Singh C and Vyas D. Use of *G. lucidum* extract to elevate the resistance in chickpea against the *Fusarium oxysporum* f. sp. *ciceris*. *Arch. of Phytopathol. and Plant Protection.* 2023; 17:1-20.
 26. [^]Raseta M, Miskovic J, Capelja E, Zapora E, Petrovic Fabijan A, Knezevic P and Karaman M. Do *Ganoderma* Species Represent Novel Sources of Phenolic Based Antimicrobial Agents? *Molecules.* 2023; 28(7):3264.
 27. [^]Patel DK, Dutta SD, Ganguly K, Cho SJ and Lim KT. Mushroom-derived bioactive molecules as immunotherapeutic agents: A review. *Molecules* 2021; 26: 1359.

28. [^]Constantin M, Raut I, Suica-Bunghuez R, Firinca C, Radu N, Gurban A-M, Preda S, Alexandrescu E, Doni M and Jecu L. G. *lucidum*-Mediated Green Synthesis of Silver Nanoparticles with Antimicrobial Activity. *Materials*. 2023; 16(12):4261.
29. [^]Flores-Rabago KM, Rivera-Mendoza D, Vilchis-Nestor AR, Juarez-Moreno K and Castro-Longoria E. Antibacterial Activity of Biosynthesized Copper Oxide Nanoparticles (CuONPs) Using *Ganoderma sessile*. *Antibiotics*. 2023; 12(8):1251.
30. [^]Sudheer S, Bai RG, Muthoosamy K, Tuvikene R, Gupta VK and Manickam S. Bio sustainable production of nanoparticles via mycogenesis for biotechnological applications: A critical review. *Environ. Res.* 2022; 204: 111963.
31. [^]Sedefoglu N, Zalaoglu Y, Bozok F. Green synthesized ZnO nanoparticles using *G. lucidum*: Characterization and in vitro nano fertilizer effects. *J. of Alloys and Compounds*. 2022; 918:165695.
32. [^]Karaca B, Çoleri Cihan A, Akata I and Altuner EM. (2020). Anti-Biofilm and Antimicrobial Activities of Five Edible and Medicinal Macrofungi Samples on Some Biofilm Producing Multi Drug Resistant *Enterococcus* Strains. *Turkish J. of Agri. Food Sci. and Technol.* 2020; 8(1): 69–80.
33. ^{a, b, c}Alizadeh S, Seyedalipour B, Shafieyan S, Kheime A, Mohammadi P and Aghdami N. Copper nanoparticles promote rapid wound healing in acute full thickness defect via acceleration of skin cell migration, proliferation and neovascularization. *Biochem. Biophys. Res. Commun.* 2019; 517: 684-90.
34. ^{a, b}Vijayakumar V, Samal SK, Mohanty S and Nayak SK. Recent advancements in biopolymer and metal nanoparticle-based materials in diabetic wound healing management. *Int. J. Biol. Macromol.* 2019; 122: 137-48.
35. [^]Gong T, Yan R, Kang J and Chen R. 2019. Chemical components of *Ganoderma*. In Lin, Z. and Yang, B. (eds.) *Ganoderma and Health. Adv. in Experim. Med. and Biol.* Springer, Singapore. 2019; 1181: 59-106.
36. ^{a, b}Robles-Hernandez L, Salas-Salazar NA, Gonzalez-Franco AC. Purification and characterization of antibacterial activity against phytopathogenic bacteria in culture fluids from *G. lucidum*. *Molecules*. 2021; 26(18):5553.
37. [^]Sim CSF, Cheow YL, Ng SL and Ting ASY. Antifungal activities of metal-tolerant endophytes against *G. boninense* under the influence of metal stress. *Biol. Control*. 2019; 130: 9-17.
38. [^]Wang X and Wu J. Modulating effect of fatty acids and sterols on skin aging. *J. Funct. Foods*. 2019; 57: 135-40.
39. ^{a, b, c}Costa TM, Lenzi J, Paganelli CJ, Filho HH, Alberton MD, Tavares LB, de Oliveira D. Liposoluble compounds from *Ganoderma lipsiense* grown on solid red rice medium with antiparasitic and antibacterial properties. *Biotechnol. and Appl. Biochem.* 2020; 67(2):180-5.
40. [^]Noverita N and Ritchie YH. Antibacterial Activities of Ethanol Extracts Fruit Bodies of *G. Lucidum* and *R. Microphorus* Against *E. Coli* and *S.aureus*. *J. of Trop. Biodiver.* 2020;1(1):35-46.
41. [^]Wahba AE, El-Sayed AK, El-Fallal AA and Soliman EM. New antimalarial lanostane triterpenes from a new isolate of Egyptian *Ganoderma* species. *Med. Chem. Res.* 2019; 28: 2246-51.
42. ^{a, b}Serrano-Márquez L, Trigós A, Couttolenc A, Padrón JM, Shnyreva AV, Mendoza G. Antiproliferative and antibacterial activity of extracts of *Ganoderma* strains grown in vitro. *Food Sci. and Biotechnol.* 2021;30(5):711-21.
43. [^]Costa TM, Kaufmann V, Paganelli CJ, Siebert DA, Micke GA, Alberton MD, Tavares LBB and De Oliveira D. Kinetic identification of phenolic compounds and potential production of caffeic acid by *Ganoderma lipsiense* in solid-state fermentation. *Bioprocess Biosyst. Eng.* 2019; 42: 1325-32.

44. [^]Lu SY, Shi QQ, Peng XR, Zhou L, Li XN and Qiu MH. Isolation of benzolactones, Ganodumones A–F from *G. lucidum* and their antibacterial activities. *Bioorganic Chemistry*. 2020; 98:103723.
45. ^{a, b}Sande E, Baraza DL, Ooko S and Nyongesa PK. (2020). Isolation, characterization and antibacterial activity of ergosta-5, 7, 22-triene-3 β , 14 α – Diol (22Z) from Kenyan *G. lucidum*. *Asian J. Appl. Chem. Res.* 2020; 5:48-57.
46. ^{a, b}Al-jumaili MMO, Al-dulaimi FKY and Ajeel MA. The role of *G. lucidum* uptake on some hematological and immunological response in patients with coronavirus (COVID-19). *Syst. Rev. Pharm.* 2020; 11: 5.
47. [^]Cor Andrejč D, Knez Z and Knez Marevci M. Antioxidant, antibacterial, antitumor, antifungal, antiviral, anti-inflammatory and neuro-protective activity of *G. lucidum*: An overview. *Frontiers in Pharmacol.* 2022;13:934982.
48. [^]Nour El-Dein MM, El-Fallal AA, Ahmed El-Sayed KA and El-Esseily SR. Antimicrobial Activities of *G. mbrekobenum* Strain EGDA (Agaricomycetes) from Egypt. *Int. J. of Med. Mushroom.* 2023; 25(9): 31-41.
49. ^{a, b, c}Nguyen TT, Nguyen TT, Nguyen HD, Nguyen TK, Pham PT, Tran LT, Tran LT and Tran MH. Integrating in Silico and In Vitro studies to screen Anti-Staphylococcus aureus activity from Vietnamese *G. multiplicatum* and *G. sinense*. *Nat. Prod. Commu.* 2023; (4):1934578X231167289.
50. [^]Hu H, Liu Y, Liang X, Li X, Mo W, Xie Y, Zhang Z and Wu Q. Artificial cultivation anti-tumor activity of *G. mbrekobenum*. *Sains Malaysiana.* 2021;50(3):723-33.
51. [^]Qin FY, Yan YM, Tu ZC and Cheng YX. (\pm) Cochlearoids N–P: three pairs of phenolic meroterpenoids from the fungus *G. cochlear* and their bioactivities. *J. of Asian nat. products res.* 2019;21(6):542-50.
52. ^{a, b}Pimentel de Araujo F, Monaco M, Del Grosso M, Pirolo M, Visca P and Pantosti A. *S. aureus* clones causing osteomyelitis: a literature review (2000–2020). *J. Glob. Antimicrob. Resist.* 2021; 26:29-36.
53. [^]MaheshC, Galappaththi A, Patabendige NM, Premarathne BM, Hapuarachchi KK, Tibpromma S, Dai DQ, Suwannarach N, Rapior S, Karunarathna SC. A Review of *Ganoderma* Triterpenoids and Their Bioactivities. *Biomole.* 2022; 13(1): 24.
54. [^]Hu J, Li GF, Xu FM, Li Q, Lv T, Peng TF, Yin S and Gong W. Antibacterial lanostane triterpenoids from *G. tsugae*. *J. of Asian Nat. Products Res.* 2023; 4:1-7.
55. [^]Chan YS and Chong KP. Antimicrobial activity and metabolite analysis of *G. boninense* fruiting body. *J. Pure Appl. Microbiol.* 2020; 14: 114.
56. [^]Espinosa-García VV, Mendoza G, Shnyreva AV, Padrón JM and Trigos Á. Biological Activities of Different Strains of the Genus *Ganoderma* spp. (Agaricomycetes), from Veracruz, Mexico. *Int.J. of Med. Mushrooms.* 2021; 23(2): 67-77.
57. [^]Li LF, Liu HB, Zhang QW, Li ZP, Wong TL, Fung HY, Han QB. Comprehensive comparison of polysaccharides from *G. lucidum* and *G. sinense*: Chemical, antitumor, immunomodulating and gut-microbiota modulatory properties. *Sci. Rep.* 2018; 8: 112.
58. ^{a, b}He J, Luo ZL, Tang SM, Li YJ, Li SH, Su HY. Phylogenetic analyses and morphological characters reveal two new species of *Ganoderma* from Yunnan province, China. *MycKeys.* 2021; 84: 141-162.
59. [^]Faturrahman F, Sukiman S, Suryadi BF, Sarkono S and Hidayati E. Comparison of antimicrobial activities of ethanol extract from three species of *ganoderma* original lombok island. 2020; 1-12.
60. [^]Do Dat T, Viet ND, Dat NM, My PL, Thinh DB, Thy LT, Khang PT, Hai ND, Nam HM, Phong MT and Hieu NH. Characterization and bioactivities of silver nanoparticles green synthesized from Vietnamese *G. lucidum*. *Surfaces and*

Interfaces. 2021; 27:101453.

61. ^{a, b, c}Saravanan A, Senthil Kumar P, Karishma S, VoDVN, Jeevanantham S, Yaashikaa PR and George CS. A review on biosynthesis of metal nanoparticles and its environmental applications. *Chemosphere*. 2021; 264(2): 128580.
62. [^]Ahmad MF. *Ganoderma lucidum*: A rational pharmacological approach to surmount cancer. *J. Ethnopharmacol*. 2020; 260: 113047.
63. [^]Wu YL, Han F, Luan SS, Ai R, Zhang P, Li H and Chen LX. Triterpenoids from *G. lucidum* and their potential anti-inflammatory effects. *J. of agri. and food chem*. 2019;67(18):5147-58.
64. [^]Herdiana Y, Wathoni, N, Shamsuddin S, Joni IM and Muchtaridi M. Chitosan-based nanoparticles of targeted drug delivery system in breast cancer treatment. *Polymers*. 2021; 13(11): 1717.
65. [^]Mousavi SM, Hashemi SA, Gholami A, Omidifar N, Chiang WH, Neralla VR, Yousefi K and Shokripour M. *G. lucidum* methanolic extract as a potent phytoconstituent: characterization, in-vitro antimicrobial and cytotoxic activity. *Scientific Reports*. 2023;13(1):17326.
66. ^{a, b}Bacallao-Escudero A, Guerrero-Germán P, Torres-Moreno H, Vidal-Gutiérrez M, López-Romero JC, Tejeda-Mansir A, Esqueda M and Robles-Zepeda RE. Biological Activity of *Ganoderma* Species (Agaricomycetes) from Sonoran Desert, Mexico. *Int. J. of Med. Mushrooms*. 2023;25(10): 65-76.
67. ^{a, b}Arshadi N, Nouri H and Moghimi H. Increasing the production of the bioactive compounds in medicinal mushrooms: An omics perspective. *Microb Cell Factories*. 2023;22(11):34.
68. [^]Wang L, Li JQ, Zhang J, Li ZM, Liu HG and Wang YZ. Traditional uses, chemical components and pharmacological activities of the genus *Ganoderma* P. Karst.: A review. *RSC Adv*. 2020; 10: 42084-97.
69. [^]Li XC, Liu F, Su HG, Peng C, Zhou QM, Liu J, Huang YJ, Guo L and Xiong L. Twelve undescribed derivatives of ganoderic acid isolated from *G. luteomarginatum* and their cytotoxicity against three human cancer cell lines. *Phytochem*. 2021;183:112617.
70. [^]Ayimbila F, Siriwong S, Chaiyama V, Srihanant N and Keawsompong S. Comparative study of bio-functional profile and bioactivities of polysaccharides from *G. lucidum* and *G. neo-japonicum*. *Biocata. and Agri. Biotechnol*. 2023: 102875.
71. ^{a, b}Ahmad MF. *G. lucidum*: a rational pharmacological approach to surmount cancer. *J. of Ethnopharmacol*. 2020; 260: 113047.
72. [^]Jogaiah S, Kurjogi M, Abdelrahman M, Hanumanthappa N and Tran LS. *G. applanatum*-mediated green synthesis of silver nanoparticles: Structural characterization and in vitro and in vivo biomedical and agrochemical properties. *Arabian J. of Chem*. 2019; 12(7): 1108-20.
73. ^{a, b}Kimatu BM, Fang DL, Zhao LY and Hu Q. *Agaricus bisporus* peptide fractions confer cytoprotective ability, against hydrogen peroxide-induced oxidative stress in HepG2 and Caco-2 cells. *J. of Food Measurem. and Characteriz*. 2020; 14: 2503-19.
74. [^]Liu CY, Zhao SL, Zhu CJ, Gao Q, Bai J, Si J and Chen YX. Ergosterol ameliorates renal inflammatory responses in mice model of diabetic nephropathy. *Biomed. Pharmacotherapy*. 2020; 128: 110252.
75. [^]Hilliard A, Mendonca P and Soliman KFA. Involvement of NFκB and MAPK signaling pathways in the preventive effects of *G. lucidum* on the inflammation of BV-2 microglial cells induced by LPS. *J. of Neuroimmunol*. 2020; 345:

577269.

76. [^]Hassan F, Ni S, Becker TL, Kinstedt CM, Abdul-Samad JL, Actis LA and Kennedy MA. Evaluation of the antibacterial activity of 75 mushrooms collected in the vicinity of Oxford, Ohio (USA). *Int.J. of med. mushrooms*. 2019; 21(2):131-41.
77. [^]Elisashvili V, Asatiani MD, Khardziani T and Rai M. Natural Antimicrobials from Basidiomycota Mushrooms. *Promising Antimicrobials from Nat. Products*. 2022; 323-53.
78. [^]Sedefoglu N, Zalaoglu Y, Bozok F. Green synthesized ZnO nanoparticles using *G. lucidum*: Characterization and in vitro nanofertilizer effects. *J. of Alloys and Compounds*. 2022;9(18):165695.
79. [^]El-Borady OM, Ayat MS, Shabrawy MA and Millet P. Green synthesis of gold nanoparticles using Parsley leaves extract and their applications as an alternative catalytic, antioxidant, anticancer and antibacterial agents. *Adv. Powder Technol*. 2020; 31(10): 4390-4400.
80. [^]Singh J, Kumar S, Alok A, Upadhyay SK, Rawat M, Tsang DCW, Bolan N and Kim KH. The potential of green synthesized zinc oxide nanoparticles as nutrient source for plant growth. *J. of Cleaner Production*.2019; 214: 1061-70.
81. [^]Raseta M, Miskovic J, Capelja E, Zapora E, Petrovic Fabijan A, Knezevic P, Karaman M. Do Ganoderma Species Represent Novel Sources of Phenolic Based Antimicrobial Agents? *Molecules*. 2023;28(7):3264.
82. ^{a, b}Thapa R, Maharjan R, Tamang P, Gautam P, Adhikari R and Maharjan S. Antimicrobial assessment and phytochemical screening of medicinal plants and *G. lucidum*. *Int. J. of Appl.Sci.Biotechnol*. 2022; 10: 228-36.
83. [^]Suansia A and John P. Antimicrobial and Antioxidant properties of medicinal mushroom *Ganoderma P. Karst*. *GSCBiol. and Pharmaceu.Sci*. 2021; 17: 106-12.
84. [^]Chan YS and Chong KP. Bioactive compounds of *G. boninense* inhibited methicillin-resistant *S. aureus* growth by affecting their cell membrane permeability and integrity. *Molecules*. 2022; 27(3): 838.
85. ^{a, b}Huang WC, Chang MS, Huang SY, Tsai CJ, Kuo PH, Chang HW, Kao MC. Chinese herbal medicine *G. tsugae* displays potential anti-cancer efficacy on metastatic prostate cancer cells. *Int. J. Mol. Sci*. 2019; 20: 4418.
86. [^]Muniroh MS, Nusaibah SA, Vadamalai G and Siddique Y. Proficiency of biocontrol agents as plant growth promoters and hydrolytic enzyme producers in *G. boninense* infected oil palm seedling. *Curr. Plant Biol*. 2019; 20: 100116.
87. [^]Kamal A, Batool M, Saba M and Albsher G. Wild Mushroom (*Ganoderma multipileum*) as Biosource for Zinc oxide Nanoparticles: From Synthesis to Enhance Biological Applications. *Authorea*. 2023; 1-13.
88. ^{a, b, c}SaqibS, FaryadS, AfridiMI, ArshadB, YounasM, NaeemM, Bimetallic assembled silver nanoparticles impregnated in *Aspergillus fumigatus* extract damage the bacterial membrane surface and release cellular contents. *Coatings*. 2022; 12(10): 1505.
89. [^]Devi HS, Boda MA, Shah MA, Parveen S and Wani AH. Green synthesis of iron oxide nanoparticles using *Platanus orientalis* leaf extract for antifungal activity. *Green Process. Synth*. 2019; 8(1):3 8-45.
90. [^]Alizadeh S, Seyedalipour B, Shafieyan S, Kheime A, Mohammadi P and Aghdami N. Copper nanoparticles promote rapid wound healing in acute full thickness defect via acceleration of skin cell migration, proliferation and neovascularization. *Biochem. Biophys. Res. Commun*. 2019; 5(17): 684-90.
91. [^]Peng XR, Unsicker SB, Gershenzon J and Qiu MH. Structural diversity, hypothetical biosynthesis, chemical synthesis and biological activity of *Ganoderma meroterpenoids*. *Nat. Product Reports*. 2023; 40(8): 1354-92.
92. [^]Zhao M, Tang Y, Xie J, Zhao Z and Cui H. Meroterpenoids produced by fungi: Occurrence, structural diversity,

- biological activities, and their molecular targets. *European J. of Med. Chem.* 2021; 209: 112860.
93. [^]Agarwal A, Gupta V, Yadav AN, Sain D, Rahi RK, Bera SP and Neelam D. Aspects of mushrooms and their extracts as natural antimicrobial agents: *Microbiology. J. of microbiol.Biotechnol. and food sci.* 2023;12(6): e9191.
94. [^]Robles-Hernandez L, Salas-Salazar NA and Gonzalez-Franco AC. Purification and characterization of antibacterial activity against phytopathogenic bacteria in culture fluids from *G. lucidum*. *Molecules.* 2021; 26(18): 1-12.
95. [^]Tamilselvan N and Rajesh K. Antimicrobial efficacy of medicinal mushroom *G. lucidum*. *Int. J. of Trend in Sci. Res. and Development.* 2019; 3:1798-1800.
96. [^]Serrano-Márquez L, Trigos A, Couttolenc A, Padrón JM, Shnyreva AV and Mendoza G. Antiproliferative and antibacterial activity of extracts of *Ganoderma* strains grown in vitro. *Food Sci. and Biotechnol.* 2021;30(5):711-21.
97. [^]Couttolenc A, Diaz-Porras A, Espinoza C, Medina ME and Trigos A. On the primary and secondary antioxidant activity from hydroxy-methylcoumarins: experimental and theoretical studies. *J. of Physical Organic Chem.* 2020;33: e4025.
98. [^]Murillo-Rabago EI, Vilchis-Nestor AR, Juarez-Moreno K, Garcia-Marin LE, Quester K and Castro-Longoria E. Optimized synthesis of small and stable silver nanoparticles using intracellular and extracellular components of fungi: An alternative for bacterial inhibition. *Antibiotics.* 2022;11(6):800.
99. ^{a, b}Bahrulolum H, Nooraei S, Javanshir N, Tarrahimofrad H and Mirbagheri VS. Green synthesis of metal nanoparticles using microorganisms and their application in the agrifood sector. *J. Nanobiotechnol.* 2021; 19: 86.
100. [^]Nguyen VP, Le Trung H, Nguyen TH, Hoang D and Tran TH. Synthesis of biogenic silver nanoparticles with eco-friendly processes using *G. lucidum* extract and evaluation of their theranostic applications. *J. Nanomater.* 2021; 20(21): 6135920.
101. [^]Rajeshkumar S, Menon S, Kumar SV, Tambuwala MM, Bakshi, HA, Mehta M, Satija S, Gupta G, Chellappan DK, Thangavelu L, Dua K. (2019). Antibacterial and antioxidant potential of biosynthesized copper nanoparticles mediated through *Cissus arnotiana* plant extract. *Journal of Photochemistry and Photobiology B: Biology*, 197: 111531.