

[Open Peer Review on Qeios](#)

The dual energy supply of eukaryotic cells

János Hunyady

Funding: No specific funding was received for this work.

Potential competing interests: No potential competing interests to declare.

Abstract

The regeneration of tissue damage is possible because our cells have a dual-energy supply system and can ensure tissue regeneration without O₂. The publication summarizes the defining elements of the structures responsible for energy and energy-carrier transformation (SET), specifically, the hypothetical ADP-producing unit, the SET of anaerobic glycolysis (SET-AG), and the SET of oxidative phosphorylation (SET-OP). SET-AG is responsible for the anaerobic fermentation, while SET-OP is for the aerobic oxidative phosphorylation. The importance of Hypoxia Inducible Factor (HIF)-1 α in tissue regeneration is also discussed.

János Hunyady MD¹

¹*University of Debrecen, Medical Center Department of Dermatology, 4032 Debrecen, Nagyerdei krt 98, Hungary. E-mail: hunyadi@med.unideb.hu; drhunyadij@gmail.com*

Keywords: eukaryotic cell; HIF; cell energetic; tissue regeneration; Fe-S cluster.

Energy conversion

Gasoline or petrol, used as a fuel in spark-ignited internal petrol engines, must be made by fractional distillation of petroleum. Similarly, glucose must be transformed into Adenosine Triphosphates (ATP) to get a usable energy-carrier for living organisms.

The human body comprises eukaryotic cells, so it is essential to know the properties of their energy supply. This communication summarizes the evolution of eukaryote cells and their energy supply path — the dual energetic stock results in the possibility of the regeneration of tissue damage.

Energy and energy-carrier transformation, ATP synthesis.

Glycolysis and oxidative phosphorylation are autonomous mechanisms. It is well known that the energy supply of cells is

provided by glycolysis which occurs in the cytosol of cells. During glycolysis, glucose breaks down into pyruvate and energy; a total of 2 ATP is derived in the process: $\text{Glucose} + 2 \text{NAD}^+ + 2 \text{ADP} + 2 \text{Pi} \rightarrow 2 \text{Pyruvate} + 2 \text{NADH} + 2 \text{H}^+ + 2 \text{ATP} + 2 \text{H}_2\text{O}$. The specific form of glucose used in glycolysis is glucose 6-phosphate. Under aerobic conditions, Pyruvate derived from glucose will enter the mitochondria to undergo oxidative phosphorylation. Anaerobic conditions result in Pyruvate staying in the cytoplasm and being converted to lactate by the enzyme lactate dehydrogenase. [1][2] Energy is liberated in the cells during energy transformation. At the same time, ATP, one new energy-carrier molecule, will be created.

We suppose that a hypothetical structure is responsible for ADP production. Based on this hypothesis, it is proposed that glucose, NH_3 , uric acid, and H_2PO_4^- will result in the formation of ATP. In addition, ribose, the part of the adenosine + CO_2 , will be created from the D-Glucose during the process.

Energy and energy-carrier transformation is realized in unique permanent structures such as Structure for Energy Transformation (SET). Starter unit (SU), Adenosine Diphosphate Producing Unit (ADP-PU), D-glucose-6phosphate Producing Unit (G6p-PU and Pi- Producing Unit (Pi-PU), are the basic units of SETs.

The SET of anaerobic glycolysis (SET-AG) is responsible for the anaerobic fermentation, while the SET of oxidative phosphorylation (SET-OP) is for the aerobic oxidative phosphorylation.

The development of eukaryotic cells

There was no O_2 in Earth's atmosphere more than three billion years ago. At that time, the possibility of the formation of life was already ensured. The earliest cells to produce oxygen were the cyanobacteria (blue-green algae), which evolved oxygen via photosynthesis. The appearance of O_2 in the atmosphere caused the first environmental disaster, as the ancient fermenting microorganisms did not have sufficient defense capacity against the highly destructive O_2 .

According to Lynn Margilus' hypothesis, an ancient cell entered into symbiosis with a cell that could defend itself against the dangerous effects of O_2 (Illustration 1). In addition, the modern cell produced an order of magnitude more energy with the help of O_2 . The contemporary organelle is now known as a mitochondrion. [3]

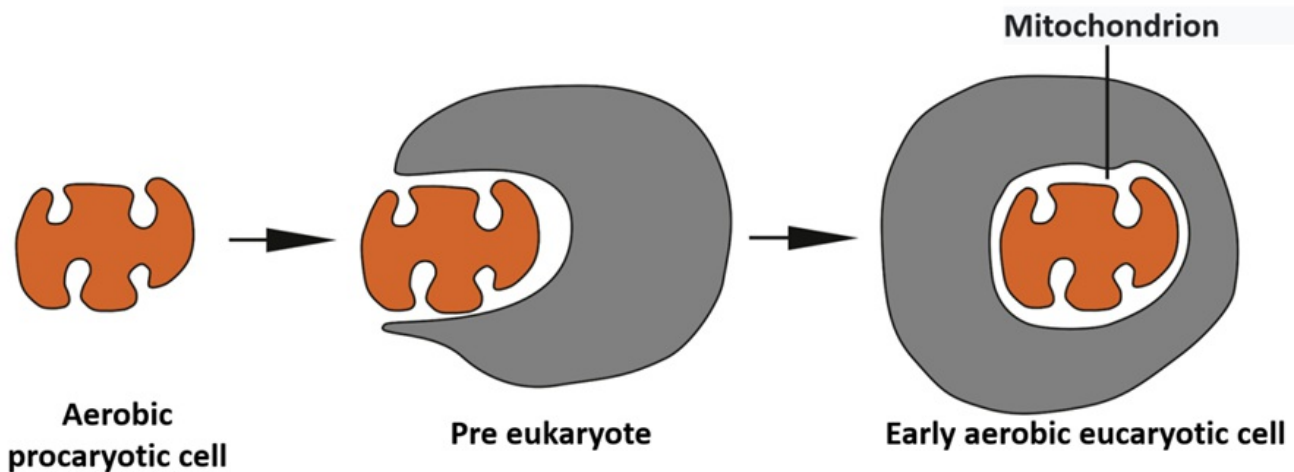


Illustration 1. The prokaryotic cell, which developed features of an early mitochondrion (defense system against reactive oxidative species and aerobic energy production), fuses with pre-eukaryote to give rise to early aerobic eukaryotic cell. ^[3]

The Evidence Supporting the Endosymbiotic Conception: ^{[3][4]}

- Mitochondria are capable of division, and their dimensions and form are like today's bacteria.
- They have their DNA, which is identical in structure to the DNA of prokaryotes.
- They have a protein-synthesizing system, similar to prokaryotes.

The advantages of symbiosis are significantly more energy, protection against free radicals, and the regeneration ability of organisms. ^{[3][4]}

The peroxisome

Peroxisome is a membrane-bound oxidative organelle, a type of micro-body, found in the cytoplasm of virtually all eukaryotic cells. ^{[5][6][7]} They perform critical roles in lipid metabolism and the conversion of reactive oxygen species. They also contain approximately 10% of the total activity of two enzymes (Glucose-6-phosphate dehydrogenase and 6-Phosphogluconate dehydrogenase) in the pentose phosphate pathway, ^[8] which is essential for energy metabolism. ^[9] Key players in peroxisome division are conserved in animals, plants, and fungi, and key fission components are shared with mitochondria. ^[10]

The electron transport chain

An electron transport chain (ETC) is a series of protein complexes and other molecules that transfer electrons from electron donors to electron acceptors via redox reactions (both reduction and oxidation co-occurring) and couples this electron transfer with the transfer of protons (H^+ ions) across a membrane. The electrons transferred to the ETC involve four multi-subunit large enzyme complexes and two mobile electron carriers. Many of the enzymes in the electron

transport chain are membrane-bound. ^{[11][12]}

Regulation by the HIF system

In the case of anoxia, the oxidative phosphorylation enzymes are inactive. In this situation, the HIF system helps realize regeneration after tissue damage.

The Hypoxia-Induced Factor (HIF)-1 α subunit is continuously synthesized and degraded under normoxic conditions, while it accumulates rapidly following exposure to low oxygen tensions. Thus due to the lack of O₂ caused by injury or any cause, the hydrolysis of the HIF-1 α is annulled. ^{[13][14]} Subsequently, the HIF system influences about 200 genes, converting cells to ancient energetic. As a result, the circulation will be restored with the help of newly formed blood vessels. After that, increasing tissue O₂ will hydrolyse the HIF-1 α ; thus, the cells will return to the mitochondrial oxidative phosphorylation. ^{[14][15]}

The dual energy supply of eukaryotic cells

Eukaryotic cells have two genetic stock as mitochondria contain their own. Accordingly, our cells must have two structures to ensure energy and energy-carrier transformation as well. SET-AG (belonging to the ancestral cell) and SET-OP (belonging to the mitochondria). The operational activity of these structures can be determined by the amount of ATP produced. In an oxygenated environment, SET-AG will not function, as the ATP produced by the mitochondria significantly exceeds the capacity of SET-AG, resulting in the shutdown of its activity.

In an anoxic or hypoxic environment, mitochondria stop working. At the same time, there is no hydrolysis of HIF-1 α , which will result in the activation of the SET-AG.

The importance of Fe-S clusters

Several Fe-S clusters [e.g., 2Fe-2S, 4Fe-4S, P-cluster of nitrogenase 8Fe-7S (cys-S)₆] are known. They play an essential role in maintaining life by ensuring continuous electron transfer. In the central part of the 2Fe-2S cluster, two irons are bonded to two sulphurs. The two irons in the 2Fe-2S cluster can bind four more sulphurs. The iron of Fe-S clusters is Fe²⁺ (deoxy, Fe II) or Fe³⁺ (oxy, Fe III) forms. The iron's oxidation state influences the iron's binding affinity to oxygen and sulphur. In the case of Fe III, it binds the oxygen, while in the case of Fe II, the sulphur bind is preferred.

Fe II modification to Fe III results in the possibility of binding oxygen-containing molecules, such as H₂PO₄, NHO, uric acid (UA), or aminated UA. Then, in an additional step, Fe III returns to Fe II. In the Fe₂-S₂ cluster, it results in four O²⁻ productions.

Other Fe-S clusters might have similar nature. Thus, the 8Fe-7S P-cluster of nitrogenase has six Cys-S structures^[16].

Thus it might produce six O^{2-} (illustration 2).

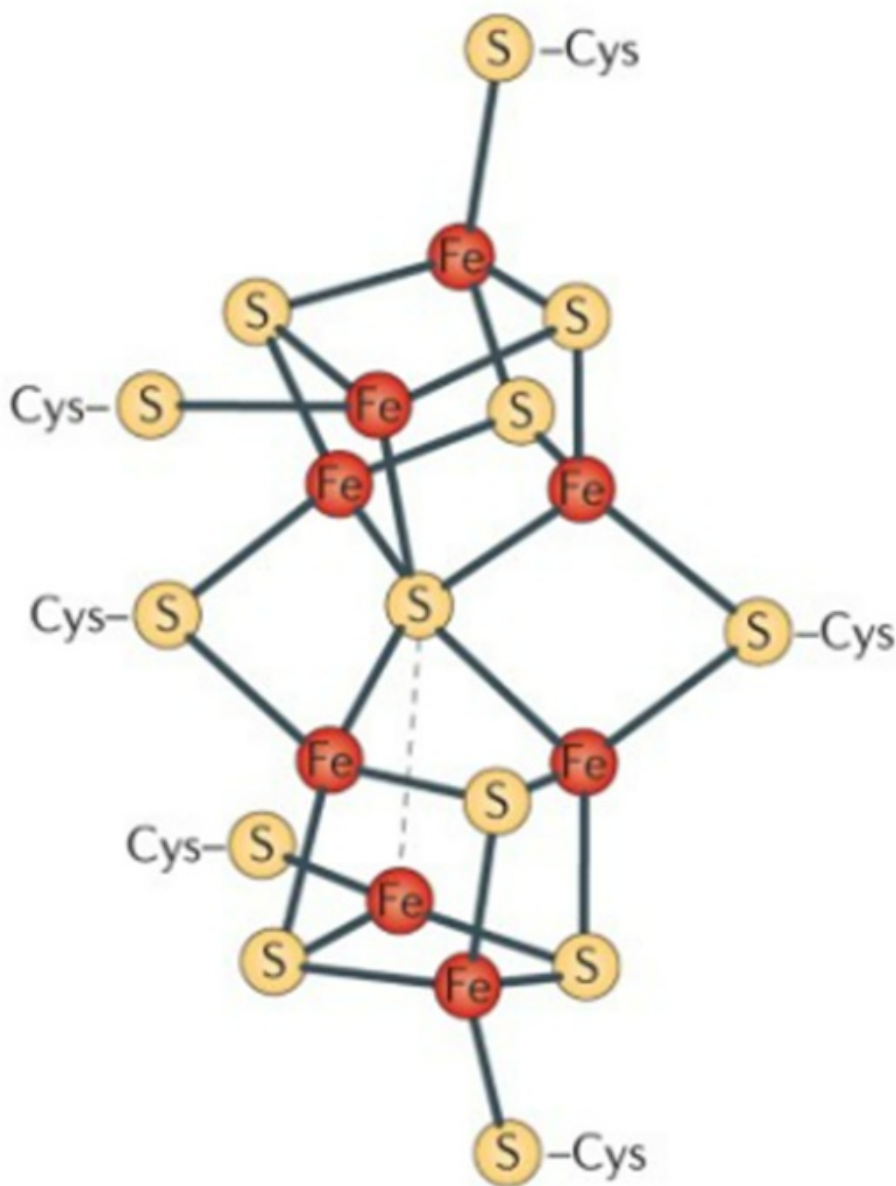


Illustration 2. 8Fe-7S (cys-S)₆ P-cluster of nitrogenase ^[16].

The functional importance of Fe-S clusters' s cys-S components.

The cys-S components of the Fe-S clusters (R-SCH₂CH (NH₂) CO₂H) contain one sulfur atom, one carboxamide, and one OH (Illustration 3)

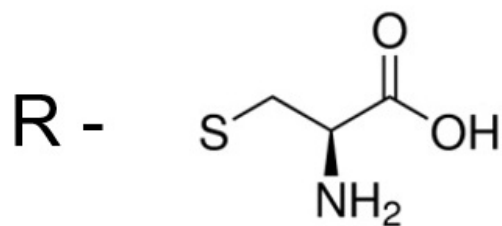


Illustration 3. The structure of cys-S.

NH₂ part of the cys-S

The NH₂ part of the structure might bind D glucose (Illustration 4) or L ascorbic acid molecules. Circle 1 indicates the binding of NH₂ to the oxygen. Circle 2 demonstrates the change of sulfur atoms to oxygen by the two OH of the AA (Illustration 5).

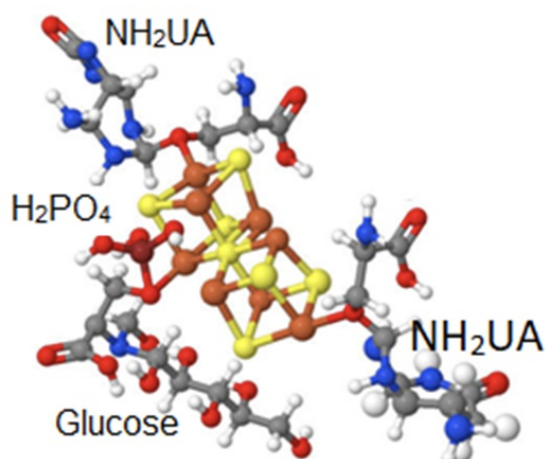


Illustration 4. Fe₈-S₇(cys-S)₆, glucose, H₂PO₄, two NH₂uric acids.(only three cys-S are illustrated).

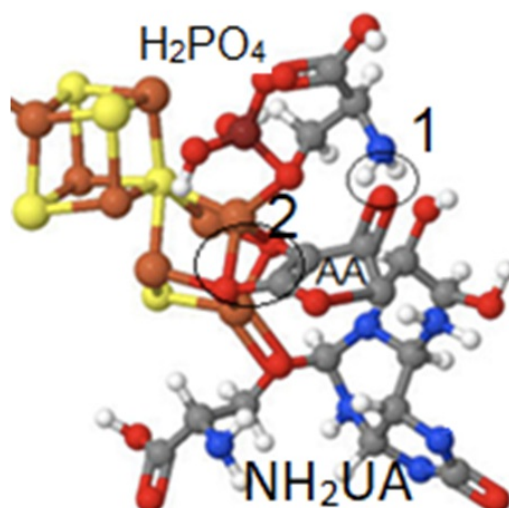


Illustration 5. Fe8-S7(cys-S)₆, H₂PO₄, Ascorbic acid (AA), NH₂ uric acid. (only three cys-S are illustrated).



Carboxyl part of the cys-S

The C=O part might bind the adenine of ATP (Illustration 6). Circle 1 indicates the binding of NH₂ to the oxygen, while circle 2 demonstrates the change of sulfur atoms to oxygen by the two OH of the ribose belonging to ATP.

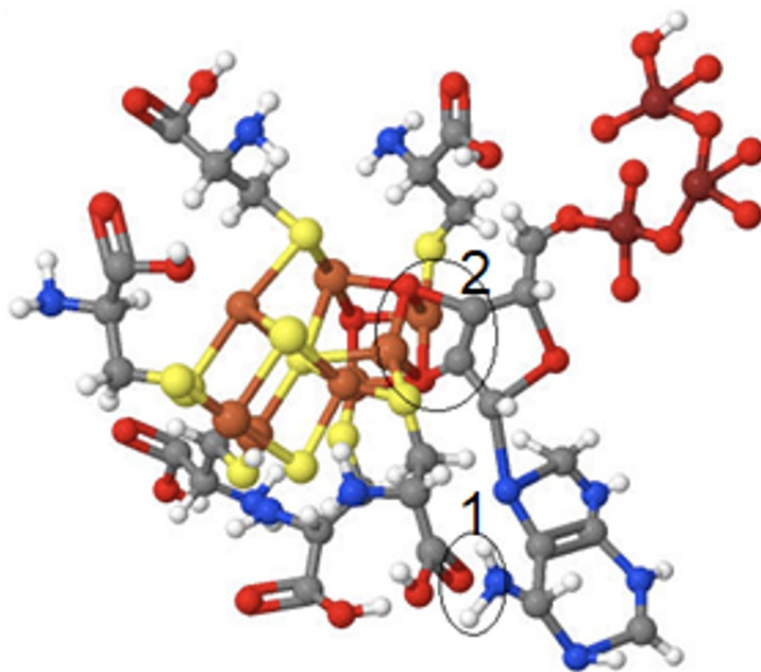


Illustration 6. Fe8-S7 (cys-S)₆, ATP



NH₂ – Carboxyl connection between the cys-S chains

Fe-S clusters might create one continuous ETC realised by the cys-S parts of the clusters. The NH₂ and the Carboxyl parts of the cys-S offer the possibility of continuous chain creation, as demonstrated in Illustration 7.

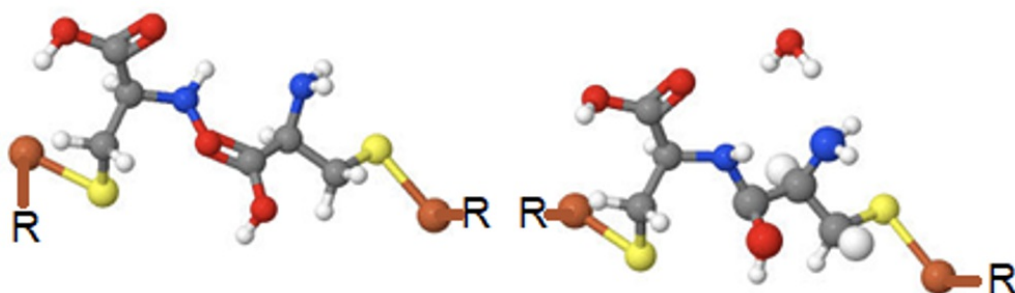
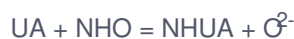
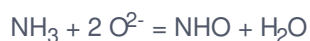


Illustration 7. Two cys-S chains are connected with the help of O^{2-}

Transformation of the source molecules.

NH_3 – NHO, Uric Acid – NHUA transformation

Two O^{2-} transforms NH_3 to $NHO + H_2O$ while one uric acid (UA) will be aminated.



D-glucose

Phosphorylation

Phosphorylation of the D-glucose is created in the D-glucose 6-phosphate Producing Unit (G6P-PU).

Glucose – ribose – Pyruvate - acetic acid transformation

Ribose + CO_2 is created from D-glucose during the transformation.



Ribose + adenine = **adenosine**

Ribose is transformed into Pyruvate and acetic acid.



$H_2PO_4^-$

Dihydrogen phosphate will be transformed into PO_3^{3-} (Pi) + two H^+ + O^{2-} in the Pi-PU and the ADP-PU.

Vitamin C and ATP are the initiators of energy transformation

Kinga Linowiecka et al. stated that ascorbic acid (AA) is an oxidative stress sensor and a gene expression regulator. In addition, they pointed out that the change of AA to DHA regulates the modulation of the iron's electron state in Fe^{2+} dependent dioxygenases (Illustration 8). [17] Two H^+ are liberated during the AA – DHA transformation.

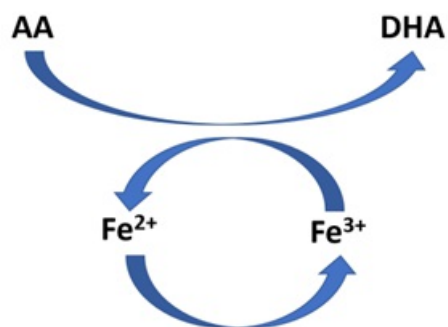


Illustration 8. Vitamin C's role in Fe²⁺ - Fe³⁺ transformation. Abbreviations: AA: ascorbic acid; DHA: dehydroascorbic acid

This change might be valid for the Fe atoms of the Fe-S clusters. The reaction results in a sulphur-oxygen exchange, creating four O²⁻ in the Fe₂-S₂ cluster.

We suppose that a similar reaction might occur by the two OH of the ribose part of the ATP initiating the Fe-S cluster.

ATP synthase

The binding change mechanism of ATP synthase involves the active site of a β subunit's cycling between three states. [18] In the "open" state, ADP and phosphate enter ATP synthase; in Illustration 15, this is shown in white. The enzyme then changes shape and forces these molecules together, with the active site in the resulting "tight" state (shown in yellow) binding the newly produced ATP molecule. Finally, the active site cycles to the loose state (red) and will be ready for the next cycle of ATP production. [18]

Structures for energy and energy-carrier transformation

SETs are places of energy and energy-carrier transformation. They contain permanent structures where the arriving molecules are converted to energy, new energy-carrier molecules (ATP), and CO₂, while the cell's membrane potential is also realized.

All SETs are built up by Starting Unit (SU), D-glucose 6 Phosphate Producing Unit (G6P-PU), PQ³⁻ (Pi)-Producing Unit (Pi-PU), ADP-PU, and ATP-synthase. Primary molecules that arrive at the structure will be transformed into new energy-carrier molecules, CO₂ and energy, while the membrane potential is also realized.

The transformation is completed in an electron transfer structure.

Four 2Fe-2S, one 3Fe-3S, and seven 4Fe-4S clusters offer the proper function of the complex, as described by Austin et

al. [19], Illustration 9.

We suppose that Electron Transfer Chain (ETC) contains one 4Fe-4S cluster and four 8Fe-7S clusters of nitrogenase instead of the seven 4Fe-4S clusters. The four 8Fe-7S clusters of nitrogenase are the determining part of the ADP Producing Unit (ADP-PU). The remaining 4Fe-4S cluster might be responsible for the D-Glucose 6-phosphate production (G6P-PU), while the four 2Fe-2S might create the Pi producing Unit (Pi-PU)

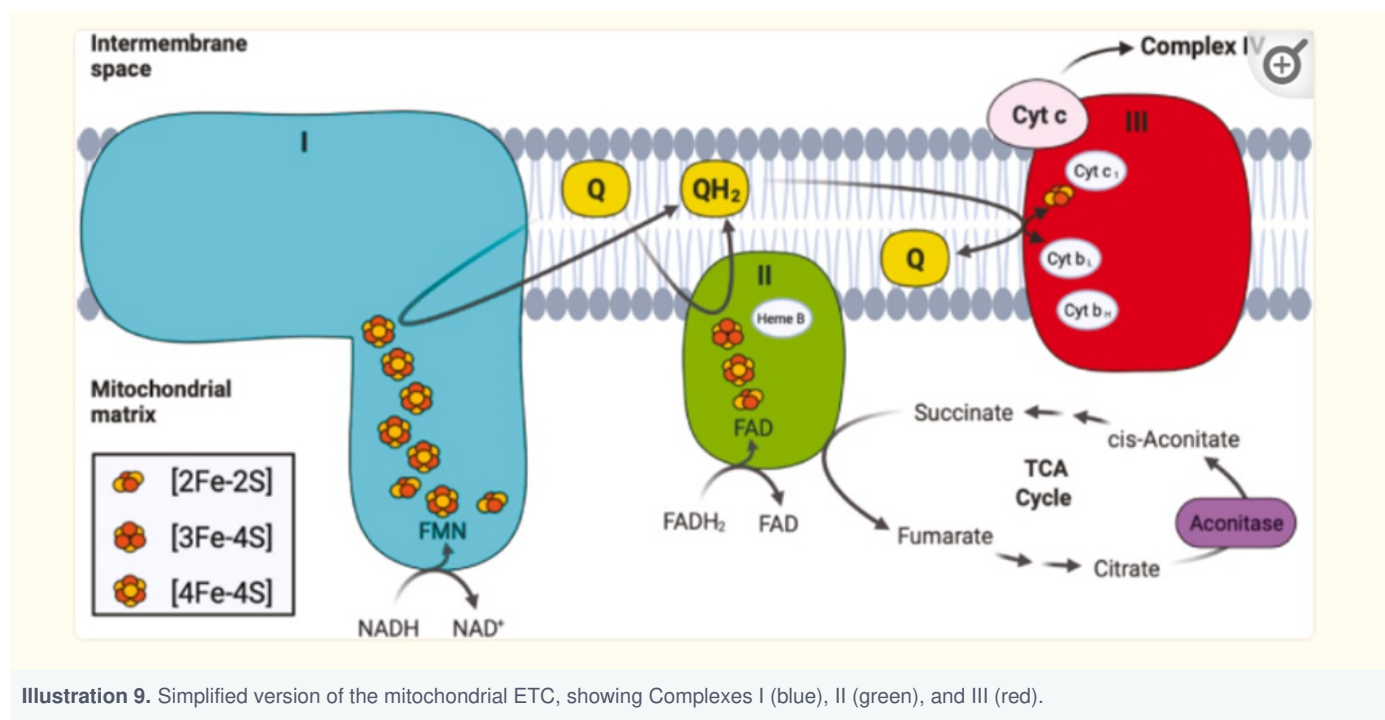


Illustration 9. Simplified version of the mitochondrial ETC, showing Complexes I (blue), II (green), and III (red).

The cys-S parts of the clusters connect Fe-S clusters and the units. The 3Fe-3S cluster might be responsible for starting the reaction and for the connection of the three specialized units (Illustration 10).

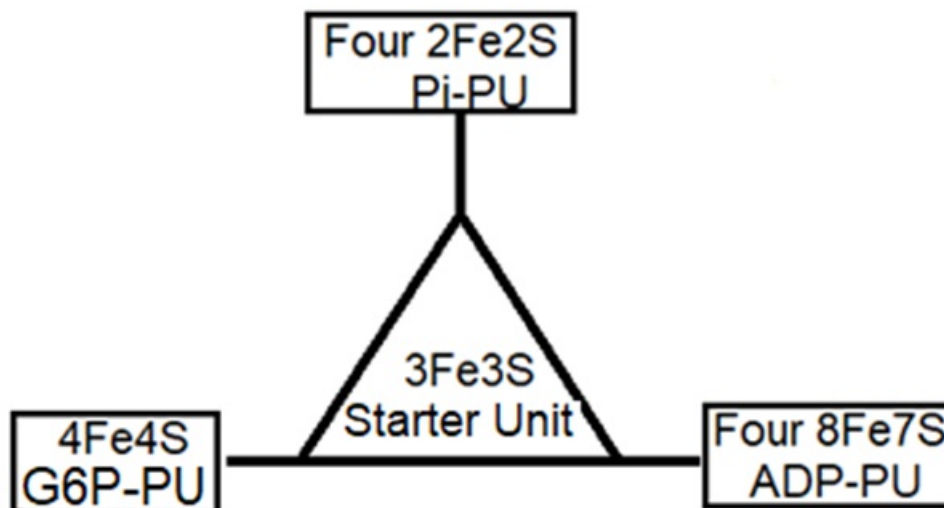


Illustration 10. The 3Fe-3S (Starter Unit) connects the three specialised units: the Pi producing Unit (Pi-PU), the D-glucose-6 phosphate Producing Unit (G6p-PU) and the ADP Producing Unit (ADP-PU).

Starter Unit (SU)

Molecule of the permanent structure:

One 3Fe-3S cluster.

Source molecules:

Three H_2PO_4^-

Products:

Three PO_3^{3-} (Pi) and three O^{2-}

The three O^{2-} atoms are responsible for the realisation of the bound between the SU and the other three units (Illustration 7).

Initiation of the unit and the ETC

One Ascorbic acid is responsible for the initiation of the structure (Illustration 11).

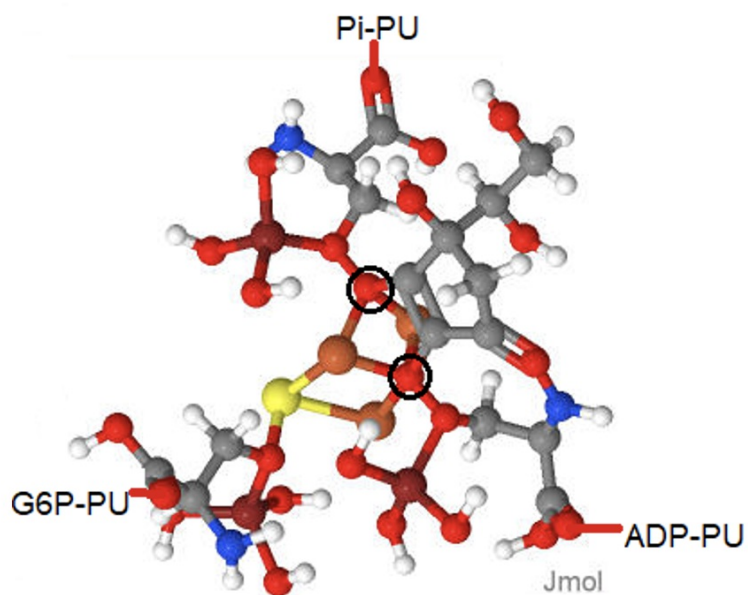


Illustration 11. Starter Unit. Two OH of one vitamin C (circled) are activating the cluster.

Abbreviations: G6P-PU: D-Glucose 6-phosphate Producing Unit; Pi-PU: PO_3^{3-} (Pi) Producing Unit; ADP-PU: ADP Producing Unit.



PO₃³⁻ (Pi) Producing Unit (Pi-PU)

Molecules of the permanent structure:

Four 2Fe-2S clusters.

Source molecules:

4 x [Acetic acid + four H_2PO_4]

Products:

4 x [four Pi + 8 H^+ + 2 CO_2 + energy].

Initiation of the unit

Four ATP are responsible for the initiation of the structure.

D-glucose-6-phosphate Producing Unit (G6P-PU)

Molecule of the permanent structure:

One 4Fe-4S cluster.

Source molecules:

Four D-Glucose + four H_2PO_4

Products:

Four D-Glucose 6-phosphate

Initiation of the unit

Two ATP are responsible for the initiation of the structure.

Adenosine diphosphate and NHO producing unit (ADP-PU)

The basic unit of SET-AG and SET-OP is the ADP-PU. In addition, ATP synthase is also required to generate ATP.

Molecules of the permanent structure:

Four 8Fe-7S clusters, one Flavin, and one nicotinamide molecule

Source molecules:

Four UA, four NH_2 -UAs, four NH_3 , four NHO, eight H_2PO_4^- , four D-glucose, and four D-glucose 6-phosphate, (Illustration 12). The unit's proper function depends on determined structure proteins and specific enzymes.

The four NH_2 -UA and eight H_2PO_4^- molecules create the tetra adenine octo phosphate ring, where four 8Fe-7S P-cluster of nitrogenase clusters connect the molecules.

Products:

4 ADP + 8 CO_2 + 16 H^+ + 4 ribose + 4 Pi + energy

Initiation of the unit

Energy investment: the initiation of the four Fe8-S7 (cys-S)₆ P-clusters is realized by four AA and 4 x 3 ATP molecules resulting 12 ADP.

The mechanism of S – O exchange might be similar to the processes of 2Fe-2S as described above.

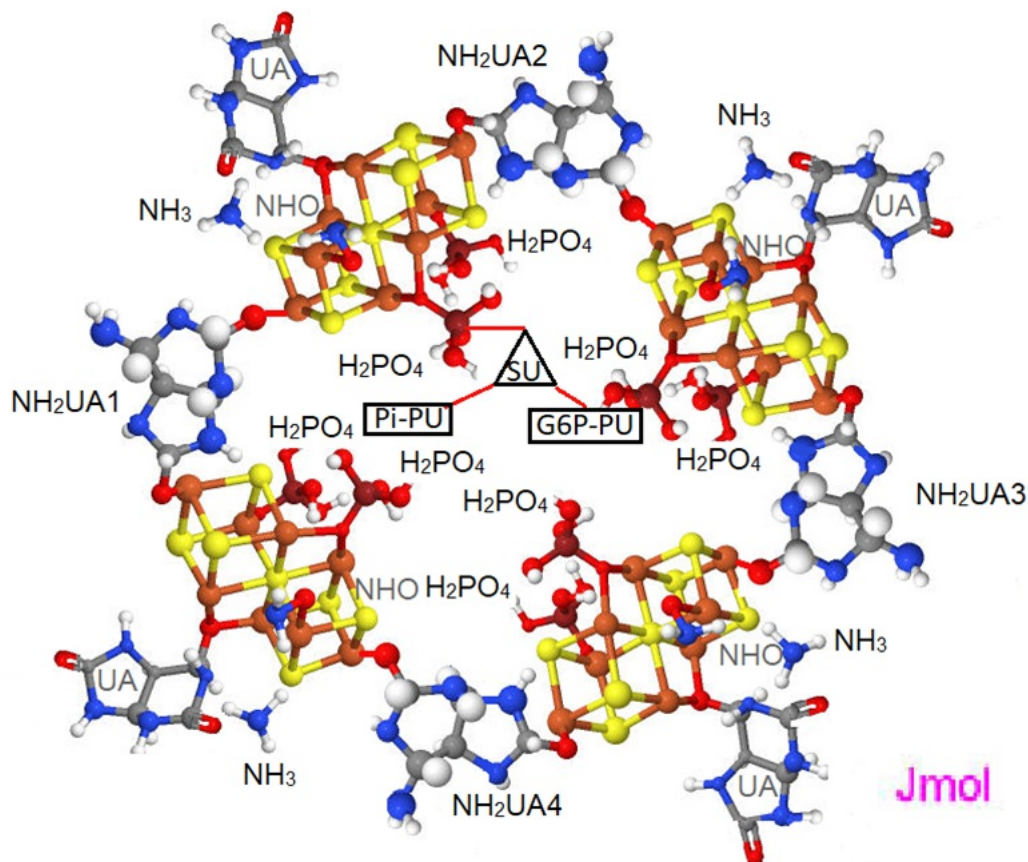


Illustration 12. Adenosine diphosphate producing unit. Four Fe8S7 (cys-S)₆, four UA, four NH₂-UA, four D-glucose, four D-glucose 6 phosphate, four NHO, four NH₃ and eight H₂PO₄⁻ in the ADP-PU.

Abbreviations: UA: Uric Acid, SU: Starting Unit, Pi-PU: Pi Producing Unit, G6P-PU: Glucose 6-Phosphate Producing Unit.



Glucose molecules are not presented.

Sulfur – Oxygen change

The affinity of FeII to OH is bigger than to S.

FeII binding OH will have three electrons.

FeIII will become FeII after the liberation of the hydrogen (Table I).

Table I. Electron transfer in Fe-S clusters

Fe II	Fe II	Fe III	Fe III	Fe II
S	OH	OH	O	S
Liberation of hydrogen H ⁺				

The four Fe₈-S₇ (cys-S)₆ clusters of the ADP-PU have 4 x 6 cys-S parts. Thus they offer places for 24 oxygen-containing molecules as eight H₂PO₄⁻, four NHO, four UA, and eight oxygen of four NH-UA molecules (Illustrations 12 and 13).

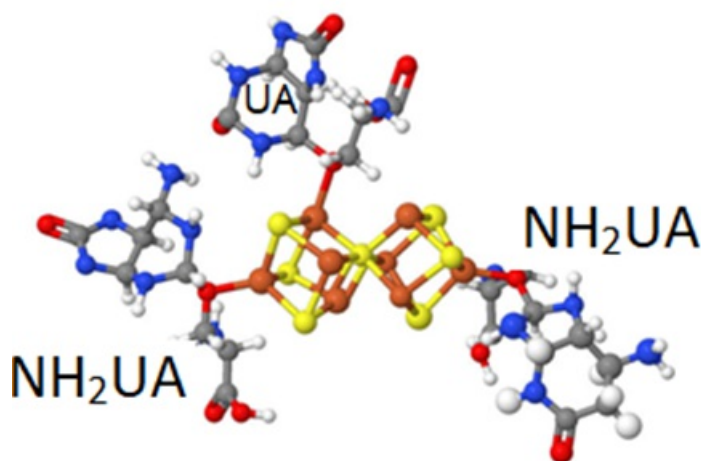


Illustration 13. 8Fe-7S (cys-S)₆, uric acid (UA), and two NH₂uric acids (only three cys-S are illustrated).



8Fe-7S (cys-S)₆

In each of the six cys-S three ATP one AA and two D-glucose molecules are situated. The remaining three NH₂ and three C=O structures offer connecting points for the stabilization of the complex structure of the SET. Illustration 14a demonstrates two 8Fe-7S clusters bounded by two cys-S. Illustration 14b shows one NH₂UA molecule attached to the structure.

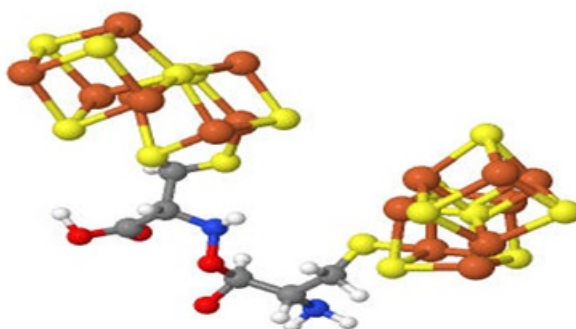


Illustration 14a. Two Fe8-S7 clusters bounded by two cys-S waiting for the NH₂UA.

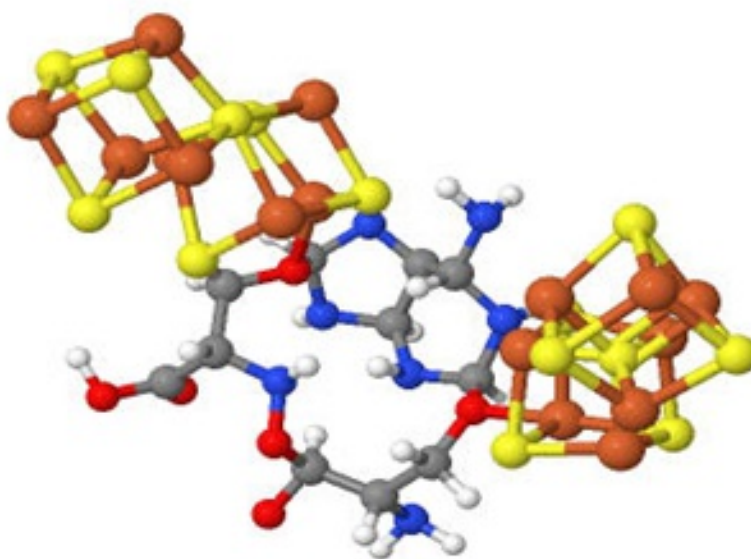


Illustration 14b. NH₂UA molecule attached to the structure.



The change of Fe II to Fe III

The two OH of AA on the lactone rings (Illustration 5) and the two OH of the ATP's ribose (Illustration 6) change the nature of the Fe atoms from Fe II to Fe III.

SET is initiated by AA, and ATP. Their ratio determines the activation. A high intracellular AA level increases the activity of SET, and a high ATP level decreases it, as it takes the place of the AA.

The production activity and sensitivity threshold of SET-AG and SET-OP are different. The ATP level reached by SET-OP inhibits the activity of SET-AG.

AA given intravenously initiates SET-AG. During this, ADP-PU produces 24 O^- .

- 8 O^{2-} mediates the $\text{NH}_3 - \text{NHO}$ transformation

$$4 \text{NH}_3 + 8 \text{O}^{2-} = 4 \text{NHO} + 4 \text{H}_2\text{O}$$
- 16 O^{2-} produces 8 CO_2 by oxidizing the eight carbon atoms of 8 D-glucose 6-phosphate resulting in eight ribose.

Suppose the activity has started, but there is no glucose in the cell. In that case, O^- destroys the tumor cell after the exhaustion of the caspase protection system.

Tumor cells consume 200 times more sugar than normal cells and their ROS activity is lower compared to the normal cells, causing their sensitivity for AA. [20]

Structure of SET-AG and SET-OP

Structure for anaerobe glycolysis

SET-AG consists of three ADP-Pus (ADP-PU-A, ADP-PU-B, and ADP-PU-C) and three ATP synthase. These structures work together in a synchronized way. When ADP-PU releases the ADP, the ATP synthase is in the open phase, ready to accept it. Furthermore, when ADP-PU-A is in the open state, ADP-PU-B is in the tight, and ADP-PU-C is in the loose state. This synchronization ensures continuous membrane potential and ATP formation (Illustration 15). [21][22]

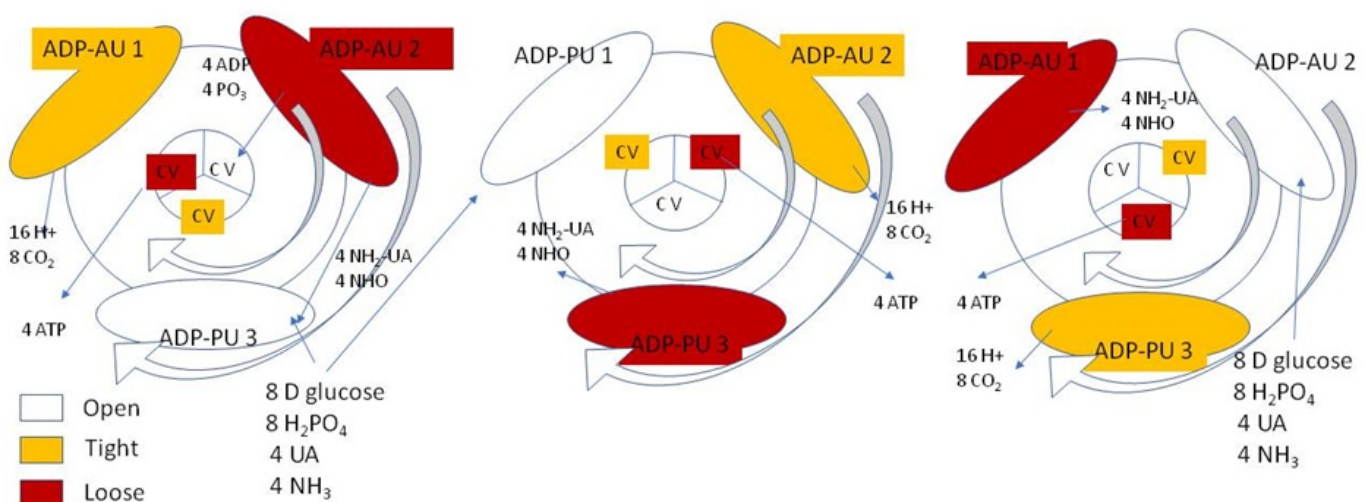


Illustration 15. The synchronised function of three ADP-Pus (I, II, III) and three ATP synthase

The four UAs with four NHO molecules forms four aminated UAs + four H₂O, while the four aminated UAs produce four adenine molecules.

In the transformation process, eight ribose molecules are created from eight D-glucose molecules.

Four ribose with four UA-originated adenine molecules forms four adenosines. In an O₂-free environment, four lactates are formed from the four Pyruvates, while in an oxygenated environment, 4x3 CO₂ + twelve H₂O molecules + energy are realized through oxidative phosphorylation. During the energy transformation, the carbon atoms of the four citric acids are converted into eight CO₂. We suppose that SET-AG has three, and SET-OP has nine ADP-PU^{[23][24]}

Structure for oxidative phosphorylation

The SET-OP consists of three SET-AG - (3x3) ADP-PU. It also contains one Pyruvate dehydrogenase complex (PDC) and three high molecular weight cytochromes (Hmc).^{[23][24]}

Location of the SET-AG and SET-OP in the cells

Austin et al. suggest that Complex 1 is in the mitochondrial membrane hanging in the mitochondrial matrix^[19]. In our hypothesis, ADP-PU is built up by four Fe₈-S₇ (cys-S)₆ (P-cluster of nitrogenase) instead of the six 4Fe-4S clusters suggested by Austin et al. The unit's proper function needs two 2Fe-2S clusters, complex II, III, and IV (Illustration 16).

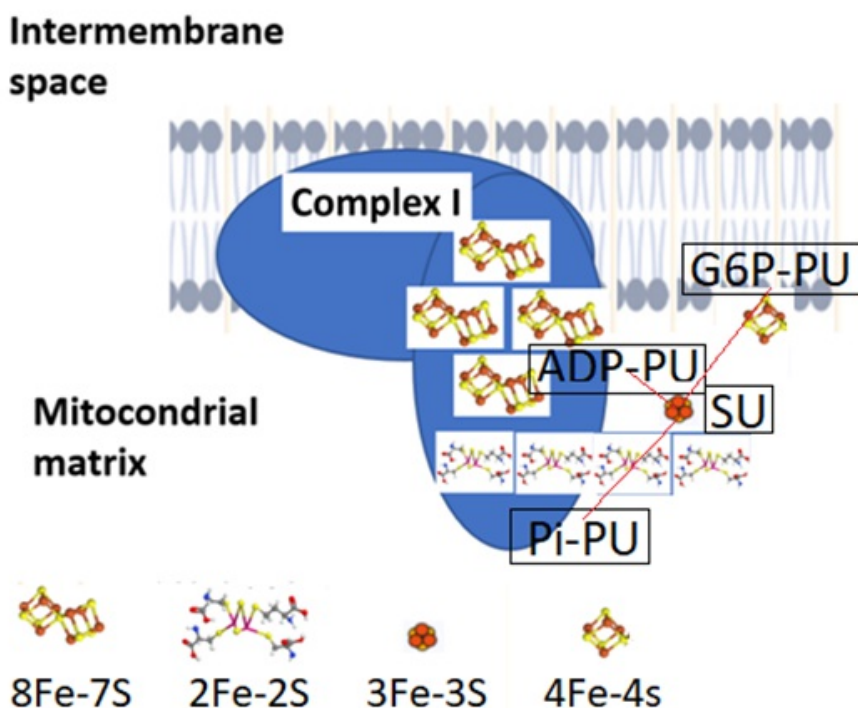


Illustration16. Four 8Fe-7S (cys-S)₆ clusters form the ADP-PU of the SET-OP, in the intermembrane space hanging in the mitochondrial matrix.

Abbreviations: ADP-PU: ADP Producing Unit, G6P-PU: Glucose 6-Phosphate Producing Unit, SU: Starting Unit, Pi-PU: Pi Producing Unit.



We assume that SET-AG is located in the peroxisomes or near the cytoplasmic membrane, while SET-OP is in the intermembrane space hanging in the mitochondrial matrix.

The hypothetical way of the energy transformation

The efficiency of ADP-producing unit

After energy investment, energy is produced in the SET. In addition, new ATP molecules are created, and the realization of the membrane potential becomes possible. At the end of the process, the ADP molecules formed during the energy investment are transformed back into ATP using the energy produced.

The hypothetical way of the energy transformation is summarised in Table II.

Table II. The hypothetical way of the energy transformation.

	Source molecules	Product						Starting molecule		
		O ²⁻	CO ₂ + energy	H ⁺		Pi	O ²⁻	ATP	AA	
I Starter Unit: one 3Fe-3S	3 H ₂ PO ₄	3					3		1	
II Pi-PU: four 2Fe-2S (cys-S)₄	16 H ₂ PO ₄ 4 acetic acid	16	8	32		16		3	1	
III G6P-PU: One 4Fe-4S	4 H ₂ PO ₄ 4 D-glucose	4			4 glucose 6 phosphate		4	1	1	
IV ADP-PU: four 8Fe-7S (cys-S)₆		24								
A. 4NH ₃ , 4 NHO 4UA + 4 NHO = 4 NHUA + 4 O ²⁻ 4 NH ₃ + 8 O ²⁻ = 4 NHO + 4 H ₂ O B. 4 NHUA + 8 H ₂ PO ₄ + 4 D-glucose + 4 D-glucose 6-phosphate = 4 ADP + 4 PO ₃ ³⁻ + 4 ribose + 8 CO ₂ + 16 H ⁺	4 UA 4 NHUA 4 NH ₃ 4 NHO 8 H ₂ PO ₄ 4 Glucose 4 glucose 6-phosphate				4 NHO 4 NHUA 4 ribose 4 ADP	4 for ATP		12	4	
V ATP synthase										
4 ADP + 4 PO ₄ ³⁻ = 4 ATP									4 ATP	

The role of HIF in the control of tissue regeneration

Cells will become viable in a hypoxic environment with the help of the HIF system, which ensures adaptation to a hypoxic environment. HIF-1 α combines with HIF-1 beta to modify the activity of about 200 genes. The most significant changes are: [\[6\]\[7\]\[25\]](#)

1. Genetic changes results in the reactivation of SET-AG.
2. Due to the low energetic efficiency of SET-AG, the appropriate energy supply of the cell can be realized only by about two hundred times more glucose. Therefore, the number of glucose transporters in the cells increases.
3. The sensitivity to apoptosis decreases.

4. Induction of neovascularization.
5. Induction of the formation of pluripotent cells.

As a result of these changes, the cells survive in the hypoxic environment and ensure the realization of tissue regeneration and neovascularisation (Illustration 17). [6][7][25]

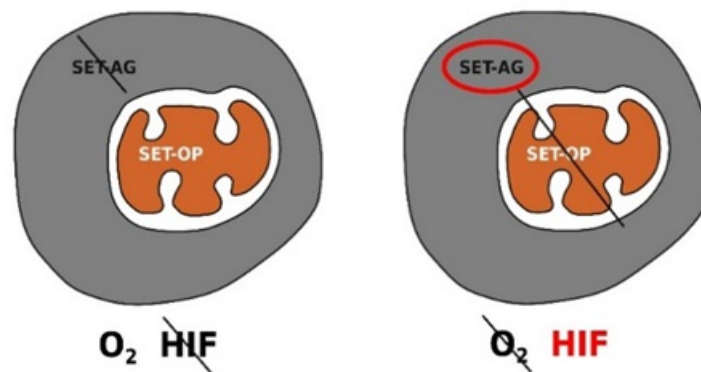


Illustration 17. The HIF system is the detector and organizer of the oxygenated and O₂-free environment.

Abbreviations: Structure for Energy Transformation of Aerobic Glycolysis:
 SET-AG; Structure for Energy Transformation of Oxidative Phosphorylation:
 SET-OP; HIF: Hypoxia-Inducible Factor.

SET-AG is always present in the cell but does not function in normoxic conditions. The Fe-S clusters of SET-AG may be in their determined place.

The hypothetical structures responsible for the energy and energy-carrier transformation must be much more complicated than described here. Their proper functions must depend on the transport and stabilizing-proteins and of many enzymes and enzyme cofactors.

Conflicting Interests

The author declared no potential conflicts of interest concerning the publication of this article.

References

1. [^] Alfarouk K, Verduzco D, Rauch C, Muddathir A, Adil H, Elhassan G, Ibrahim M, David-Polo-Orozco J, Cardone R, Reshkin S, Harguindey S. Glycolysis, tumor metabolism, cancer growth, and dissemination. A new pH-based

- etiopathogenic perspective and therapeutic approach to an old cancer question. *Oncoscience* 2014; 1(12), 777-802.
DOI: 10.18632/oncoscience.109
2. ^a Chaudhry R, Varacallo M: *Biochemistry, Glycolysis*. <https://www.ncbi.nlm.nih.gov/books/NBK482303/?report=printable>
 3. ^{a, b, c, d} Margulis L. *Origin of Eukaryotic Cells*: Yale University Press. 1970 ISBN-10: 0300013531, ISBN-13: 978-0300013535
 4. ^{a, b} Cooper GM. *The Cell: A Molecular Approach*. 2nd edition. 2000, Bookshelf Washington, DC: ASM Press; ID: NBK9841, Sunderland, Mass.: Sinauer Associates.
 5. ^a Definition of PEROXISOME. www.merriam-webster.com. Retrieved 2019-10-30.
 6. ^{a, b, c} Islinger M, Voelkl A, Fahimi HD, Schrader M. "The peroxisome: an update on mysteries 2.0". *Histochemistry and Cell Biology*. 2018; 150 (5): 443–471. doi:10.1007/s00418-018-1722-5. PMC 6182659. PMID 30219925.
 7. ^{a, b, c} O'Connell JD, Zhao A, Ellington AD, Marcotte EM. "Dynamic reorganization of metabolic enzymes into intracellular bodies". *Annu Rev Cell Dev Biol*. 2012; 28: 89–111. doi:10.1146/annurev-cellbio-101011-155841. PMC 4089986. PMID 23057741.
 8. ^a Antonenkov, Vasily D. "Dehydrogenases of the pentose phosphate pathway in rat liver peroxisomes". *European Journal of Biochemistry*. 1989; 183 (1): 75–82. doi:10.1111/j.1432-1033.1989.tb14898.x. ISSN 0014-2956. PMID 2753047.
 9. ^a Wanders RJ, Waterham HR "Biochemistry of mammalian peroxisomes revisited". *Annual Review of Biochemistry*. 2006; 75: 295–332. doi:10.1146/annurev.biochem.74.082803.133329. PMID 16756494.
 10. ^a Schrader M, Bonekamp NA, Islinger M. Fission and proliferation of peroxisomes. *Biochim Biophys Acta*. 2012;1822(9):1343-57. doi: 10.1016/j.bbadis.2011.12.014. Epub 2011 Dec 31.
 11. ^a Lyall, Fiona (2010). "Biochemistry". *Basic Science in Obstetrics and Gynaecology*. pp. 143–171. doi:10.1016/B978-0-443-10281-3.00013-0. ISBN 978-0-443-10281-3.
 12. ^a https://en.wikipedia.org/wiki/Electron_transport_chain#Complex_I
 13. ^a Salceda S and Caro J. Hypoxia-inducible Factor 1a (HIF-1a) Protein Is Rapidly Degraded by the Ubiquitin-Proteasome System under Normoxic Conditions. *THE JOURNAL OF BIOLOGICAL CHEMISTRY* 1997; Vol. 272, No. 36, Issue of September 5, pp. 22642–22647,
 14. ^{a, b} Wenger RH, Stiehl D P, Camenisch G: *Integration of oxygen signaling at the consensus HRE*. *Sci STKE*. 2005 18;2005(306):re12. doi: 10.1126/stke.3062005re12
 15. ^a Rezvani RH, Ali N, Nissen LJ, Harfouche G, Verneuil HD. HIF-1 α in Epidermis: Oxygen Sensing, Cutaneous Angiogenesis, Cancer, and Non-Cancer Disorders. *Journal of Investigative Dermatology*, 2011; 131:1793–1805. PMID: 21633368, DOI: 10.1038/jid.2011.141
 16. ^{a, b} Stephen M. Keable SM, Zadvornyy OA, Johnson LE, Ginovska B, Rasmussen AJ, Danyal K, Eilers BJ, Prussia GA, LeVan AX, Simone Raugei S, Seefeldt LC, and Peters JW: Structural characterization of the P1 intermediate state of the P-cluster of nitrogenase *J. Biol. Chem.* (2018) 293(25) 9629 –9635.
 17. ^a Linowiecka K, Foksinski M and Brożyna AA: Vitamin C Transporters and Their Implications in Carcinogenesis. *Nutrients* 2020; 12, 3869; doi:10.3390/nu1212386
 18. ^{a, b} Nakamoto RK, Baylis Scanlon JA, Al-Shawi MK. "The rotary mechanism of the ATP synthase". *Archives of*

Biochemistry and Biophysics. 2008;476 (1): 43–50.

19. ^{a, b}Austin DR, Rachel ETB, Stephen LA, Kimberly JDS. Mitochondrial iron–sulfur clusters: Structure, function, and an emerging role in vascular biology *Redox Biology* 2021; Volume 47, November, 102164
20. [^]Hong, S.; Lee, S.; Moon, J.; Hwang, J.; Kim, D.; Ko, E.; Kim, H.; Cho, I.; Kang, J.; Kim, D.; et al. SVCT-2 in breast cancer acts as an indicator for L-ascorbate treatment. *Oncogene* 2013, 32, 1508–1517. [CrossRef] [PubMed]
21. [^]Hunyady J. The Hypothesis of the Structures for Energy Transformation in Living Cells; Vitamin C, the Spark Plug of Glycolysis. *Int. J. Mol. Sci.* 2022, 23, 4380. <https://doi.org/10.3390/ijms23084380>
22. [^]Hunyady J: The Role of Vitamin C in the Energy Supply of Cells Hypothetical Structure for Energy Transformation. *Journal of Scientific Research & Reports* 2021; 27(7): 30-44, Article no. JSRR.70812 ISSN: 2320-0227
23. ^{a, b}Gresser MJ, Myers JA, Boyer PD. "Catalytic site cooperativity of beef heart mitochondrial F1 adenosine triphosphatase. Correlations of initial velocity, bound intermediate, and oxygen exchange measurements with an alternating three-site model". *The Journal of Biological Chemistry*. 1982; 257 (20): 12030–12038. doi:10.1016/S0021-9258(18)33672-X. PMID 6214554.
24. ^{a, b}Huang L, Gu J, Schau M, Bunn H. Regulation of hypoxia-C 1alpha is mediated by an O2-dependent degradation domain via the ubiquitin-proteasome pathway. *Proc Natl Acad Sci USA* 1998; 95(14):7987-7992. <https://doi.org/10.1073/pnas.95.14.7987>.
25. ^{a, b}Grano, A.; De-Tullio, M. Ascorbic acid as a sensor of oxidative stress and a regulator of gene expression: The yin and yang of Vitamin C. *Med. Hypotheses* 2007, 69, 953–954. [CrossRef]